

1 **Chemical profile and eco-safety evaluation of essential oils and hydrolates from**  
2 ***Cistus ladanifer*, *Helichrysum italicum*, *Ocimum basilicum* and *Thymbra capitata***

3

4 Celso Afonso Ferraz<sup>1,2</sup>, Ana Catarina Sousa<sup>2,3,4\*</sup>, Débora Caramelo<sup>5</sup>, Fernanda

5 Delgado<sup>5,6,7</sup>, Ana Palmeira de Oliveira<sup>1,8</sup>, M. Ramiro Pastorinho<sup>2,9</sup>

6

7 <sup>1</sup>Health Sciences Research Centre (CICS), University of Beira Interior, Covilhã, Portugal

8 <sup>2</sup>NuESA-Health and Environment Study Unit, Faculty of Health Sciences, University of

9 Beira Interior, Covilhã, Portugal

10 <sup>3</sup>Department of Biology, **School of Sciences and Technology**, University of Évora, Évora,

11 Portugal

12 <sup>4</sup>Comprehensive Health Research Centre (CHRC), University of Évora, Évora, Portugal

13 <sup>5</sup>Plant Biotechnology Center of Beira Interior (CBPBI), Castelo Branco, Portugal.

14 <sup>6</sup>Polytechnic Institute of Castelo Branco-School of Agriculture (IPCB-ESA), Castelo

15 Branco, Portugal

16 <sup>7</sup>Research Centre for Natural Resources, Environment and Society (CERNAS-IPCB),

17 Instituto Politécnico de Castelo Branco, Portugal

18 <sup>8</sup>Labfit–Health Products Research and Development Lda, UBImedical, Covilhã, Portugal

19 <sup>9</sup>Department of Medical and Health Sciences, **School of Health and Human**

20 **Development**, University of Évora, Évora, Portugal

21 \* Corresponding author: [acsousa@uevora.pt](mailto:acsousa@uevora.pt)

## 22 **Abstract**

23 The demand for natural-based products for **industrial applications** is increasing sharply  
24 and therefore the search for new alternatives to the **plants traditionally used** is growing.  
25 These alternative plants can be an important source of bioactive compounds under a  
26 circular economy approach. Considering the potential future use of **new** plant extracts  
27 by the industry, it is necessary to assess the risk associated with their introduction into  
28 the ecosystem. This work aims to provide an insight into the volatile profiles and  
29 evaluate the potential (eco)toxic effects of essential oils (EO's) and hydrolates of four  
30 plant species, namely rockrose (*Cistus ladanifer*), curry plant (*Helichrysum italicum*),  
31 conehead thyme (*Thymbra capitata*) and basil (*Ocimum basilicum*). Chemical analysis  
32 was performed by GC-MS and acute toxicity tests were performed using the model  
33 organism *Daphnia magna*. The essential oil and the hydrolate from *H. italicum*, as well  
34 as all the other hydrolates caused no immobilization **up to** the highest concentrations  
35 tested, suggesting that all hydrolates present low to no risk towards *D. magna*. Similarly,  
36 the essential oil of *H. italicum*, presented negligible risk towards *D. magna*. For *C.*  
37 *ladanifer* and *T. capitata* essential oils, the EC<sub>50</sub> (the concentration estimated to  
38 immobilize 50 per cent of the *Daphnia*) at 48h varied from 199.7 **mg/L** and 12.05 **mg/L**,  
39 respectively. The essential oil from *C. ladanifer* was mainly characterised by  
40 monoterpene hydrocarbons, while the *H. italicum* was richer in sesquiterpene  
41 hydrocarbons. Both essential oil and hydrolate from *T. capitata* contained exclusively  
42 monoterpene hydrocarbons with a particularly high content of carvacrol. The higher  
43 acute toxicity of *T. capitata* essential oil can be attributed to the high amount of  
44 carvacrol **present in the distillate**. Overall, of the essential oils and hydrolates tested, all  
45 can be classified as practically non-toxic, except for *T. capitata* essential oil that,

46 according to the Globally Harmonized System of Classification and Labelling of  
47 Chemicals (GHS) of the United Nations, can be classified as moderately toxic.

48 **Keywords:** Aromatic plants, bioactive compounds, ecotoxicity, invertebrates, acute  
49 toxicity, *Daphnia magna*

50

## 51 **1. Introduction**

52 Plants have been used for centuries for their beneficial properties, and today they are  
53 an important source of bioactive organic compounds of high economic interest. Plant  
54 extracts and their purified active components have been used in several industrial  
55 applications such as the food processing, pharmaceuticals and in cosmetics (Rafinska et  
56 al., 2019). In the food industry, these compounds are mainly used for their antioxidant  
57 activity to increase the shelf life of food products as well as new functional foods that  
58 aim to promote health and decrease the probability of developing certain types of  
59 diseases (Granato et al., 2017). In the pharmaceutical industry, plant extracts are used,  
60 for example, in skincare products (Harhaun et al., 2020), in biomedicine for wound care,  
61 and healing (Renu et al., 2019) and in a wide variety of products for their antimicrobial  
62 properties. In fact, the use of plant extracts with antimicrobial activity is gaining  
63 importance, as they could be used to replace antibiotics, an extremely important aspect  
64 considering the growing phenomena of antimicrobials' resistance (Aleksic Sabo and  
65 Knezevic, 2019). Different extracts can be obtained from plants such as essential oils,  
66 and hydrolates. Essential oils are defined as the product obtained by distillation of a  
67 plant or any of its parts, or by a mechanical process (without the application of heat)  
68 from the epicarp of Citrus fruits. During the procedure of obtaining essential oils,

69 hydrolates can also be **attained** as a by-product of the process. A hydrolate is the distilled  
70 water that remains after the distillation process and is usually rich in water-soluble  
71 components of the essential oil (Hamdi et al., 2017; ISO, 2013).

72 Due to the growing interest of **industry and the general public** in natural ingredients,  
73 alternative and still largely **unexploited** plants are being studied to be used by the  
74 cosmetics and pharma industries. The use of these alternative species is particularly  
75 important for the valorisation of endogenous resources that are still largely unexplored,  
76 including, for example, *Cistus ladanifer* (L.), *Helichrysum italicum* (Roth) G. Don fil.,  
77 *Ocimum basilicum* (L.), and *Thymbra capitata* (L.).

78 *Cistus ladanifer* is an evergreen woody shrub part of the Cistaceae family (Papaefthimiou  
79 et al., 2014). It is known by the common name rockrose and it is very abundant in the  
80 wild areas of the western Mediterranean region (Spain, Portugal, south of France, and  
81 north of Morocco) (Frazão et al., 2018). It has been used in the perfumery industry due  
82 to a particular extract, the “labdanum”, used as a fixative, and in the cosmetic industry  
83 in the form of essential oil (Barrajón-Catalán et al., 2016; Zidane et al., 2013). Recent  
84 studies have shown relevant properties of its essential oil and extracts (e.g., aqueous,  
85 hydroalcoholic, acetone:water) as antimicrobial, antioxidant, cytotoxic, anti-  
86 inflammatory, and anti-nociceptive, phytotoxic, **and insecticidal** activity (Raimundo et  
87 al., 2018). As with *C. ladanifer*, *Helichrysum italicum* (Asteraceae) extracts present a  
88 wide variety of important properties and a large variety of extracts can be prepared from  
89 this plant, with resulting products differing in their chemical composition. *H. italicum*,  
90 most commonly known as curry plant or immortelle, is also widely distributed in the  
91 Mediterranean region (Kladar et al., 2015). Extracts can be obtained from several parts

92 of the plant (leaves and flowerheads, flowers, flowering tops, and aerial parts) using  
93 different solvents (acetone, diethyl ether, ethanol, methanol, or even by supercritical  
94 CO<sub>2</sub>) while the essential oil is generally obtained from the flowers (Antunes Viegas et  
95 al., 2014). The essential oil from *H. italicum* is one of the most popular essentials oils  
96 being used in cosmetics, particularly in skin regeneration and anti-age treatments, and  
97 also in soaps and perfumes due to its characteristic scent (Sarkic and Stappen, 2018).

98 *Thymbra capitata* (Lamiaceae) known generally as conehead thyme, is an important  
99 aromatic plant and its essential oils are rich in the phenolic monoterpene carvacol. The  
100 essential oil can be used to inhibit *Pseudomonas aeruginosa* biofilm formations which  
101 are often associated with multidrug-resistant infections (Qaralleh, 2019). A reduction in  
102 biofilm biomass and metabolic activity of *Candida albicans* has also been reported  
103 (Palmeira-de-Oliveira et al., 2012). This essential oil also exhibits strong antibacterial and  
104 antioxidant activity *in vitro* (Anastasiou et al., 2020) and it displays a high potential to be  
105 used as a natural preservative in emulsions (Neves et al., 2017).

106 *Ocimum basilicum* is another example of an aromatic plant from the Lamiaceae family  
107 used commercially in many countries, with particular importance in the food industry  
108 (Açıkgöz, 2020). It has been used in traditional medicine as an antispasmodic, aromatic,  
109 digestive, carminative, galactagogue, stomachic, and tonic agent (Marwat et al., 2011).  
110 The high content of polyphenols such as flavonoids and anthocyanins are responsible  
111 for the high antioxidant properties of extracts and essential oil from this plant (Ragab  
112 and Saad-Allah, 2020).

113 Although plant-based products are generally perceived as safe, some compounds  
114 extracted from the plants may be very toxic and present toxicological effects on certain

115 organisms. There are a few reports on the toxicity of plant extract towards aquatic  
116 invertebrates, particularly *Daphnia magna* Straus. This organism, commonly known as  
117 water flea, is a primary consumer from the Cladocera order, with a short life cycle. When  
118 maintained in a laboratory under ideal conditions of food, medium, temperature, pH,  
119 and photoperiod its reproduction is parthenogenic, resulting in genetically identical  
120 female offspring (Ebert, 2005; OECD, 2004). *D. magna* is one of the recommended  
121 organisms to perform ecotoxicity tests according to the Organization for Economic Co-  
122 operation and Development (OECD) and the American Society for Testing and Materials  
123 (ASTM) (ASTM, 1997; OECD, 2004).

124 The objective of this work is to characterise the volatile profiles and evaluate the toxicity  
125 profile towards *Daphnia magna* of essential oils and hydrolates obtained from  
126 Portuguese wild and cultivated aromatic plants that can be used as cosmetic and  
127 pharmaceutical ingredients, specifically the gum rockrose (*Cistus ladanifer*), curry plant  
128 (*Helichrysum italicum*), conehead thyme (*Thymbra capitata*), and basil (*Ocimum*  
129 *basilicum*).

130

## 131 **2. Materials and Methods**

### 132 **2.1 Plants, essential oils and hydrolates preparation**

133 *Cistus ladanifer* aerial parts were collected from wild plants growing in the central-west  
134 region of Portugal. *H. italicum* aerial parts were obtained from cultivated plants in the  
135 central-north region of Portugal. *T. capitata* and *O. basilicum* were obtained from  
136 cultivation fields respectively in southern Beira Interior and the Northwestern  
137 Portuguese coast. The essential oils and the respective hydrolates of *C. ladanifer* and *H.*

138 *italicum* were purchased from the companies “Aromas do Valado” and “Planalto  
139 Dourado”, respectively. They were obtained by steam distillation. However, the details  
140 on the distillation method and apparatus used were not provided by the manufacturers.  
141 The essential oil and hydrolate from *T. capitata*, were obtained from the aerial parts of  
142 the plant (flower, stem and leaf), while for *O. basilicum* hydrolate only the leaves were  
143 used. The *T. capitata* essential oil was obtained according to the procedure described in  
144 the European Pharmacopoeia (Europe, 1997). The aerial parts of *T. capitata* were  
145 harvested at the flowering stage and subjected to hydrodistillation for 2h, using a  
146 Clevenger-type apparatus, in which the essential oil and hydrolate were obtained as the  
147 final product. The leaves of *O. basilicum* were also harvested at the flowering stage and  
148 used to obtain the hydrolate by using the same process described above. All the  
149 essential oils and hydrolates obtained were stored in dark vials at 4°C until further  
150 assays.

151

## 152 **2.2 Essential oils and hydrolates analysis by GC-MS**

153 The volatile profile of the essential oils of *C. ladanifer*, *H. italicum* and *T. capitata* were  
154 analyzed, in triplicate, by gas chromatography coupled with mass spectrometry (GC/MS  
155 SCION-SQ 456 GC, Bruker) equipped with a capillary column, HP-5MS (30m × 0.25mm ×  
156 0.25 μm). Helium was the carrier gas used with a flow rate of 1mL.min<sup>-1</sup>. The initial oven  
157 temperature was programmed to 45°C, gradually increasing 3 °C/min to 175°C, finally  
158 increasing to 300°C with a heating rate of 15°C/min, maintaining this final temperature  
159 for 10 minutes. The transfer line and the ion source were programmed at a temperature  
160 of 250°C and 220°C respectively. The extracts were analyzed from electron impact

161 ionization mass spectrometry (EI-MS) at 70eV, and the compounds were identified in  
162 scan mode with positive polarity of ions 20-300 m/z with a time of 250.0 ms. All the  
163 essential oils were injected with a volume of 1  $\mu$ L, using a split ratio of 1:350 for *C.*  
164 *ladanifer* and *T. capitata*, and 1:100 for *H. italicum*, which was previously diluted with  
165 an organic solvent. Regarding the hydrolates of *H. italicum*, *O. basilicum* and *T. capitata*,  
166 they were subjected to a liquid-liquid extraction (LLE) with an organic solvent (hexane)  
167 (Collin and Gagnon, 2016; Riani et al., 2017). The aqueous phase was separated from  
168 the organic phase, and the organic phase was injected and analyzed using the same  
169 chromatographic method as essential oils. All hydrolate samples after undergoing this  
170 process were injected with a volume of 1  $\mu$ L, in triplicate, except the hydrolate of *C.*  
171 *ladanifer* which was injected with a volume of 0.5  $\mu$ L. The split ratio was 1:50 for *H.*  
172 *italicum* and *O. basilicum*, 1:20 for *T. capitata* and 1:10 for *C. ladanifer*. The  
173 identification of the compounds was based on the retention index (RI) comparing with  
174 the RI given by the MS library (NIST 17 version 2.3) and with RI calculated from the n-  
175 alkane series standards (C7-C18 and C19-C30) that was injected under the same  
176 chromatographic conditions and with the same column as the samples of essential oils  
177 and hydrolates. The relative amount of each compound was expressed as a percentage  
178 of the relative peak area of the compound relative to the total area of the peaks  
179 identified in the samples.

180

### 181 **2.3 *Daphnia magna* culture**

182 *Daphnia magna* (clone K6) stock culture was maintained in ASTM hard water medium  
183 under continuous aeration at 20°C  $\pm$  1 and a photoperiod of 16/8h light/dark cycle. The  
184 daphnids were fed daily with a suspension of the green algae *Raphidocelis subcapitata*

185 (3.0 × 10<sup>5</sup> cells/mL) from a culture maintained in house, and the culture medium was  
186 changed every other day. Before tests, adult female daphnids were isolated in 100 mL  
187 glass beakers (1 per beaker) and maintained under the same standard conditions.  
188 Parthenogenic daphnids (<24h old) descending from the isolated adults, between the  
189 2<sup>nd</sup> and 5<sup>th</sup> brood were selected to perform the tests, while the 6<sup>th</sup> brood was used to  
190 start a new culture.

191

## 192 **2.4 Range finding tests**

193 *Cistus ladanifer* and *Helichrysum italicum* essential oils chemical composition is highly  
194 heterogeneous, and therefore, a range-finding test was performed to obtain  
195 information on the appropriate concentrations to be used in the acute toxicity test  
196 (OECD, 2004). Since there is no available data on *H. italicum* essential oil toxicity towards  
197 *D. magna*, neonates (<24h old) were also exposed to widely spaced concentrations of  
198 the test substance (1, 10, 100, 500 and 1000 mg/L). The range of concentrations used  
199 for the *T. capitata* essential oil was set according to available toxicity information of the  
200 major compound present to *D. magna*. In the range-finding tests, five organisms were  
201 used by concentration in one replicate and immobilisation was observed and recorded  
202 at the end of the test (48h). This process was continuously repeated narrowing the  
203 concentrations until appropriate test concentrations were obtained.

204

## 205 **2.5 Test solutions**

206 Due to low solubility in the culture medium (ASTM hard water), DMSO was used as a  
207 solvent for the *C. ladanifer*, *H. italicum* and *T. capitata* essential oils. For the hydrolates

208 extracts, no solvent was **necessary** as they are water-based. The test substances were  
 209 weighed and dissolved in the solvent at the concentration of 1% in ASTM hard water.  
 210 When no solvent was used, the test substance was directly dissolved in ASTM hard  
 211 water. The obtained solutions were **subjected to serial dilution in culture medium** until  
 212 reaching the working concentrations (**Table 1**). When **Dimethyl sulfoxide (DMSO)** was  
 213 used as a solvent, the highest amount of solvent in the work solutions was below 0.1%  
 214 to minimize any possible effects of the solvent on the results as recommended by the  
 215 followed guideline (OECD, 2019). **An overview of the different essential oils and**  
 216 **hydrolates tested and the concentrations used are presented in table 1.**

217

218 Table 1: Essential oils and extracts concentrations used in the acute toxicity tests.

Plant	Type	Concentrations tested (mg/L)
<i>Cistus ladanifer</i>	Aerial parts	Essential oil 50; 100; 150; 200; 400
		Hydrolate 125; 250; 500; 1000; 2000
<i>Helichrysum italicum</i>	Aerial parts	Essential oil 50; 100; 200; 400; 800
		Hydrolate 125; 250; 500; 1000; 2000
<i>Ocimum basilicum</i>	Leaves	Hydrolate 500; 1000; 2000; 4000; 8000

<i>Thymbra</i>		Essential oil	5; 7.5; 10; 25; 50
<i>capitata</i>	Aerial parts		
		Hydrolate	50; 100; 150; 200; 300; 400

---

219

## 220 **2.6 Acute toxicity tests**

221 Toxicity tests were performed according to the OECD Test No. 202: *Daphnia* sp. Acute  
 222 Immobilisation Test (OECD, 2004). Neonates (<24h) from the 2<sup>nd</sup> to 5<sup>th</sup> brood were  
 223 selected for the tests. For each test, five groups with increasing concentrations of the  
 224 essential oils or hydrolates, plus the control group (ASTM medium or DMSO 0.1% -  
 225 solvent control) were tested. For each concentration, five replicates were used with five  
 226 animals per replicate. The tests were performed in multi-well plates containing 10 ml of  
 227 test medium in each well and the animals were not fed during the test. Immobilization  
 228 of the daphnids was observed and recorded at 24 and 48h. An organism was considered  
 229 immobilized if no movement was observed within 15 seconds after gentle agitation of  
 230 the test vessels. During the experiments, daphnids were kept under a constant  
 231 temperature of 20°C ± 1°C and photoperiod of 16/8 light/dark cycle and were not fed  
 232 until the end of the test. Based on the immobilisation results obtained, the EC<sub>50</sub>  
 233 (concentration estimated to immobilise 50% of the daphnids) value was calculated using  
 234 GraphPad Prism 8 software.

235

## 236 2.7 Test validation

237 To validate the tests, the reference toxic potassium dichromate ( $K_2Cr_2O_7$ ) was used as  
238 recommended in the OECD guideline. Immobilization at 24h was registered and the  $EC_{50}$   
239 value was calculated. The  $EC_{50}$  was  $1.714 \pm 1.205 \times 10^{-5}$  mg/L which is within the 0.6 –  
240 2.1 mg/L range proposed by the International Organization for Standardization (ISO)  
241 (ISO, 2012) and recommended by the OECD guideline.

242

## 243 3. Results and Discussion

### 244 3.1 Phytochemical profile

245 The analysis of essential oils from the four selected species shows that *C. ladanifer*  
246 essential oil is mainly composed of monoterpene compounds. Similar results were  
247 obtained for the *T. capitata*, for which all major compounds belong to the class of  
248 monoterpene compounds. In contrast, *H. italicum* essential oil has a higher percentage  
249 of sesquiterpene compounds and some monoterpenes. The hydrolates of *H. italicum*  
250 and *T. capitata* display different compositions in relation to their essential oils. For *H.*  
251 *italicum*, the hydrolate sample main compound was  $\alpha$ -terpineol (30.5%), while for the  
252 essential oil  $\gamma$ -curcumene (16.1%) was the major compound. Concerning *T. capitata*, the  
253 major compound, carvacrol, was the same in both essential oil and hydrolate with an  
254 amount of 79.9% and 98.1% respectively. It is also legitimate to state that all compounds  
255 belong to the monoterpene class. The *O. basilicum* hydrolate was characterized mainly  
256 by a compound widely used in perfumes and flavours, eugenol, and it is the one that has  
257 the lowest percentage of monoterpene compounds. Very low yields were obtained for  
258 *O. basilicum* essential oil hydrodistillation process and thus only a hydrolate was

259 acquired and analysed in this work. On the other hand, the essential oil and the  
260 hydrolate of *C. ladanifer* showed some differences regarding the quantity and quality of  
261 the two extracts. In relation to essential oil, it has a large percentage of monoterpenes,  
262 in which the major compound is  $\alpha$ -pinene and in the case of hydrolate, the major  
263 compound is 4-Hydroxy-3-methylacetophenone and  $\alpha$ -pinene is not among the five  
264 compounds majority. This shows the importance of analyzing the chemical profile of the  
265 two types of extracts since they may have different compounds that could possibly have  
266 different bioactivities. All the data is shown in Table 2.

267 Table 2: Chemical composition of *C. ladanifer*, *H. italicum*, *T. capitata* and *O. basilicum* and essential oils and respective hydrolates (except for  
 268 *O. basilicum* for which no essential oil was obtained)

Compounds	<i>C. ladanifer</i>				<i>H. italicum</i>				<i>T. capitata</i>				<i>O. basilicum</i>	
	EO		Hydrolate		EO		Hydrolate		EO		Hydrolate		Hydrolate	
	RI <sup>a</sup>	Peak Area (%)	RI	Peak Area (%)	RI	Peak Area (%)	RI	Peak Area (%)	RI	Peak Area (%)	RI	Peak Area (%)	RI	Peak Area (%)
α-pinene	920	35.8			917	7.4								
Camphene	930	6.7												
β-myrcene								968	2.1					
o-cymene	1004	4.6												
ρ-cymene								1000	5.5	995	0.4			
α-terpinene								991	1.8					
1,8-cineole							1011	15.4			1002	0.4	1007	3.4
2,2,6-trimethylcyclohexanone	1014	6.7												
γ-terpinene								1037	6.1					
Linalool													1084	38.3
Camphor													1129	1.1
Endo-borneol			1156	8.4			1160	3.0						
Terpinen-4-ol							1173	2.0			1166	0.7		

δ-Terpineol				1162	6.6				
p-cymen-8-ol		1179	10.7						
α-Terpineol				1188	30.5			1182	3.1
(-)-Myrtenol		1191	11.2						
D-verbenone		1205	9.8						
Bornyl acetate	1289	4.9					-		
Carvacrol				1311	29.6	1316	79.9	1311	98.1
4-Hydroxy-3-methylacetophenone		1310	21.6						
Eugenol								1364	52.5
Nerol acetate			1379	11.5					
(-)-Italicene			1388	12.5					
γ-Curcumene			1493	16.0					
α-Curcumene			1496	10.1					
Class composition									
Monoterpene hydrocarbons		80.2	34.0	32.8	96.5		100	100	46.7
Oxygenated monoterpenes		8.4							
Sesquiterpene hydrocarbons				67.2					
Others		11.5	66.0		3.5				53.4

269 <sup>a</sup>RI: Retention index calculated by the standard n-alkane series.

270

271

### 272 3.2 Acute toxicity

273 The essential oils tested exhibited different levels of toxicity to *D. magna*. Generally,  
274 when detected, the acute toxicity of the essential oils and extracts was dose and time-  
275 dependent. The essential oil from the aerial parts of *C. ladanifer* caused immobilisation  
276 of the daphnids at high concentrations (48h EC<sub>50</sub> ~200 mg/L). The essential oil obtained  
277 from the aerial parts of *H. italicum* showed no observable toxic effects up to the highest  
278 concentration tested of 800 mg/L. The essential oil obtained from the aerial parts of *T.*  
279 *capitata* caused the highest observable toxic effects, presenting the lowest EC<sub>50</sub> value  
280 of 10.81 mg/L (95% CI 9.55 – 12.60) after 48h of exposure (Table 3).

281

282 Table 3: *Daphnia magna* acute toxicity of essential oils and extracts after 24 and 48h of  
283 exposure with the indication of the respective EC<sub>50</sub> and 95% Confidence Interval (CI).

Plant	Type	Parts used	EC <sub>50</sub> mg/L (24h)	95% (CI) mg/L	EC <sub>50</sub> mg/L (48h)	95% (CI) mg/L
<i>Cistus</i>	Essential	Aerial	201.1	*	199.7	*
<i>ladanifer</i>	oil	parts	0		0	
<i>Thymbra</i>	Essential	Aerial	12.05	11.03 –	10.81	9.55 –
<i>capitata</i>	oil	parts		13.31		12.60

284 \* Not possible to calculate

285

286 All the hydrolates tested caused no observable acute effects to *D. magna* after 48h of  
 287 exposure up to the highest concentrations tested. The hydrolates and the essential oil  
 288 that caused no immobilisation to *D. magna* are presented in Table 4 alongside the  
 289 highest concentrations tested. The maximum concentrations for which no observable  
 290 effects were observed were always above the limit to be considered toxic by  
 291 international regulations. The toxicity was ranked according to the Globally Harmonized  
 292 System for Classification and Labelling of Chemicals (GHS) proposed by the United  
 293 Nations (Table 5).

294

295 Table 4: Maximum concentrations tested of *H. italicum* essential oil and hydrolates  
 296 that caused no observable effects in *D. magna* after 48h of exposure.

Plant	Type of extract	Part(s) used	Maximum concentration tested (mg/L)
<i>Cistus ladanifer</i>	Hydrolate	Aerial parts	2000
<i>Helichrysum italicum</i>	Essential oil	Aerial parts	800
	Hydrolate		2000
<i>Ocimum basilicum</i>	Hydrolate	Leaves	8000
<i>Thymbra capitata</i>	Hydrolate	Aerial parts	400

297

298 **3.2.1 *Cistus ladanifer* essential oil and hydrolate**

299 The EC<sub>50</sub> for the *C. ladanifer* ranged between 201.1 and 199.7 mg/L at 24 and 48h,  
300 respectively. The chemical analysis of the essential oil used in this study identified 37  
301 compounds present making up 92.6% of the total composition. The most abundant  
302 compound was  $\alpha$ -pinene (35.8%) followed by camphene (6.7%), which is in accordance  
303 with other studies that evaluated the composition of essential oils obtained from *C.*  
304 *ladanifer*. Other important compounds present were 2,2,6-trimethylcyclohexanone  
305 (6.7%), bornyl acetate (4.9%), o-cymene (4.6%), D-limonene (3.3%), viridiflorol (3.5%)  
306 and pinocarveol (3.1%). Twenty eight other compounds were present at concentrations  
307 below 3% showing the high heterogeneity of compounds present in the *C. ladanifer*  
308 essential oil. The chemical composition of essential oils obtained from this plant has  
309 been intensively studied. Most of the studies report  $\alpha$ -pinene as the major compound  
310 present (Costa et al., 2007; Greche et al., 2009; Gülz et al., 1984; Mariotti et al., 1997;  
311 Robles et al., 2003; Rossi et al., 2007; Tavares et al., 2020). Camphene is usually the  
312 second most abundant compound present, although some authors reported camphene  
313 as the major compound (Zidane et al., 2013). Other compounds including trans-  
314 Pinocarveol and Viridiflorol (Gomes et al., 2005; Verdeguer et al., 2012) or 1,8-Cineole  
315 (Viuda-Martos et al., 2011) have also been reported. The acute toxicity of  $\alpha$ -pinene  
316 towards *D. magna* was reported to be in the range between 0.22 - 1.44 mg/L, whereas  
317 the acute toxicity of a mixture containing  $\alpha$ -pinene, camphene,  $\beta$ -pinene and  $\delta$ -3-carene  
318 was reported to be 4.29 mg/L in the US EPA (United States Environmental Protection  
319 Agency) database. Although  $\alpha$ -pinene is reported to be toxic to *D. magna* at low  
320 concentrations, in this study, the percentage of this compound is relatively low (~36%)  
321 and several other compounds were detected which can reduce the toxicity of the  
322 essential oil. In the European Chemicals Agency (ECHA) database, an essential oil

323 obtained from the stems and leaves of *C. ladanifer* by distillation is registered as having  
324 toxic effects to *D. magna* with EC<sub>50</sub> values ranging from 94.2 mg/L after 24h of exposure  
325 and 63.2 mg/L after 48h of exposure. It was classified as long term hazardous to the  
326 aquatic environment under the Chronic 3 category (EC<sub>50 (48h)</sub> within 10 and 100 mg/L and  
327 degradability of the test substance in the environment considered to be low (ECHA,  
328 2016)). However, no other information about the chemical composition of the essential  
329 oil or the origin of the plant material is provided by ECHA. The *C. ladanifer* essential oil  
330 used in this work presented a 48h EC<sub>50</sub> value above 100 mg/L, which cannot be  
331 considered hazardous to the environment (Table 5). These differences in the observed  
332 toxicity can be explained by the chemical variations that are common in essential oils.  
333 The chemical variability of Portuguese *C. ladanifer* essential oils was studied recently,  
334 showing differences in the chemical composition of the essential oils according to the  
335 time of the year the plant was harvested and the extraction process (steam distillation  
336 vs hydrodistillation) (Tavares et al., 2020). These normal variabilities of the chemical  
337 composition of essential oils from *C. ladanifer* can be the explanation of the different  
338 toxicity observed with the essential oil in this study and the one reported in the ECHA  
339 database. The hydrolate cause no observable effects to *D. magna* after 48h of exposure  
340 up to 2000 mg/L. As mentioned above, *C. ladanifer* essential oil and hydrolate differ  
341 slightly in their chemical profile. According to a previous study (Tavares et al., 2020) the  
342 hydrolate major compounds were trans-pinocarveol, borneol and terpinen-4-ol, which  
343 in comparison with this study only verbenone fits in the identified major compounds.  
344 On the other hand, the percentage of monoterpenes hydrocarbons was very low in  
345 relation to the other classes of compounds, which was also verified in the present work.  
346 It is important to note that in the present work, the samples were acquired commercially

347 while in the study by Tavares et al. the aerial parts of the plant were collected in Beira  
 348 Baixa in 2017 and 2018 and subjected to steam distillation. The extraction method as  
 349 well as the time of harvesting the plant are variables that influence the chemical  
 350 characterization and consequently the bioactivity analyses.

351

352 **Table 5:** Classification of essential oils and hydrolates tested according to the Globally  
 353 Harmonized System for classification and labelling of chemicals (GHS) proposed by the  
 354 United Nations. The proposed values are based on 48 hours EC<sub>50</sub> values for  
 355 crustaceans. Acute 1: ≤ 1 mg/L; 1 < Acute 2 ≤ 10 mg/L; 10 < Acute 2 ≤ 100 mg/L; Non-  
 356 toxic: >100 mg/L.

Plant	Type of extract	48h EC <sub>50</sub> mg/L	Classification
<i>Cistus ladanifer</i>	Essential oil	> 100	Non-toxic
	Hydrolate	> 100	Non-toxic
<i>Helichrysum italicum</i>	Essential oil	> 100	Non-toxic
	Hydrolate	> 100	Non-toxic
<i>Ocimum basilicum</i>	Hydrolate	> 100	Non-toxic
<i>Thymbra capitata</i>	Essential oil	10.8	Acute 3
	Hydrolate	> 100	Non-toxic

357

### 358 **3.2.2 *Helichrysum italicum* essential oil and hydrolate**

359 *Helichrysum italicum* essential oil showed no acute toxic effects to *D. magna* up to 800  
360 mg/L suggesting very low toxicity towards this organism. The chemical analysis  
361 identified 27 compounds present, which were responsible for 93.6% of the total  
362 chemical composition of the essential oil. The three most abundant compounds were  $\gamma$ -  
363 curcumene (16.1%), followed by (-)-italicene (12.6%) and neryl acetate (11.5%). It is  
364 noticeable that the compounds present in the essential oil are all in relatively low  
365 percentages. Regarding the effects of these three major compounds, there is no  
366 available data on the toxicological effects of these compounds on *D. magna* or other  
367 aquatic invertebrates. The hydrolate studied also did not cause any observable toxic  
368 effect to *D. magna* up to high concentrations (2 g.L<sup>-1</sup>). In total, 17 compounds were  
369 identified in the hydrolate, being the most abundant L- $\alpha$ -terpineol (30.6%), followed by  
370 carvacrol (29.6%) and 1,8-cineole (15.4%). There are significant chemical differences  
371 between the essential oil and the hydrolate obtained from *H. italicum*. To the authors'  
372 best knowledge, there is no available data on the literature about the chemical  
373 composition of *H. italicum* hydrolates, as this type of extract has attracted far less  
374 attention when compared with the essential oil. The high heterogeneity of compounds  
375 present in similar percentages in the essential oil and hydrolate might explain the lack  
376 of toxic effects observed to *D. magna* up to relatively high concentrations. To the  
377 authors' best knowledge, this is the first time that the toxicity towards *D. magna* of an  
378 essential oil and hydrolate from *H. italicum* are tested. The obtained results are also an  
379 important contribution for the characterization of the hydrolate which is obtained  
380 simultaneously with the essential oil, and that can be regarded as a source of bioactive  
381 compounds instead of "waste" of the distillation process.

382 The chemical composition of essential oils obtained from *Helichrysum italicum* can vary  
383 depending on the region the plant is grown, and it can present very distinct chemotypes  
384 depending on subspecies and environmental factors (e.g. soil properties). An essential  
385 oil obtained by hydrodistillation of the aerial parts of *H. italicum* from Croatia was  
386 predominantly constituted of neryl acetate (20.5%),  $\gamma$ -curcumene (14.1%) and trans- $\alpha$ -  
387 bergamotene (7.0%), and other 59 compounds were identified in lower percentages  
388 (Dzamic et al., 2019). Another essential oil obtained from the aerial parts of *H. italicum*  
389 in Montenegro showed similar chemical composition with neryl acetate and  $\gamma$ -  
390 curcumene as the two most abundant compounds (29.2% and 18.8%, respectively), but  
391 neryl propanoate was the third most abundant with 10.1% (Kladar et al., 2015).  
392 Interestingly, another study reported  $\beta$ -eudesmene,  $\beta$ -bisabolene and  $\alpha$ -pinene as the  
393 three most abundant compounds (21.6%, 19.9% and 16.9%, respectively) of an essential  
394 oil from *H. italicum* collected in Montenegro (Oliva et al., 2020). On the other hand, an  
395 essential oil from aerial parts of *H. italicum* collected in the North of Algeria showed a  
396 high diversity of compounds in small percentages, with  $\alpha$ -cedrene (13.6%),  $\alpha$ -curcumene  
397 (11.4%) and geranyl acetate (10.1%) the three most abundant (Djihane et al., 2017). In  
398 Portugal, for example, an essential oil obtained by hydrodistillation of the aerial parts of  
399 *H. italicum* subsp. *picardii* collected in the south of Portugal was rich in  $\alpha$ -pinene (53.5%)  
400 and  $\gamma$ -curcumene (27.4%) (Costa et al., 2015).

401

### 402 **3.2.3 *Thymbra capitata* essential oil and hydrolate**

403 The *Thymbra capitata* essential oil was the most toxic towards *D. magna*. The EC<sub>50</sub> values  
404 obtained varied from 12.1 mg/L after 24h of exposure and 10.8 mg/L after 48h of

405 exposure. This essential oil can be classified as acutely hazardous to the aquatic  
406 environment under the acute 3 category of the GHS (Table 5) as the 48h EC<sub>50</sub> obtained  
407 is within 10 and 100 mg/L. The toxicity observed at relatively low concentrations can be  
408 explained by the high amount of carvacrol present in the essential oil (80%). This  
409 phenolic monoterpene is registered in ECHA as being toxic to aquatic life, with a  
410 reported 48h EC<sub>50</sub> value to *D. magna* of 6.06 mg/L (95% CI 5.10 - 7.28 mg/L), 96h EC<sub>50</sub>  
411 for the zebrafish *Danio rerio* of 6.17 mg/L (95% CI 4.83 – 9.92 mg/L) and 72h EC<sub>50</sub> to the  
412 green microalgae *Raphidocelis subcapitata* of 4.05 mg/L (ECHA). This shows that  
413 carvacrol is toxic towards different aquatic organisms from microalgae to fish, and  
414 contamination of water bodies by this substance or solutions of the substance should  
415 be avoided. The observed EC<sub>50</sub> value to *D. magna* in this study follows closely the EC<sub>50</sub>  
416 value reported of carvacrol in the ECHA database. Moreover, the low biodegradability  
417 of carvacrol in the environment has led to the chronic 2 category classification, as toxic  
418 to aquatic life with long-lasting effects by the ECHA. The chemical composition of  
419 essential oils obtained from *T. capitata* are usually very similar in composition and  
420 carvacrol is consistently reported as the major compound. An essential oil obtained from  
421 the aerial parts of *T. capitata* collected in north Morocco, by hydrodistillation,  
422 presented 75.5% of carvacrol (Charfi et al., 2019). In another study, different essential  
423 oils from the aerial parts of *T. capitata* collected from 2002 to 2004 in both flowering  
424 and fruiting stages in the Badajoz area, Spain, consistently showed high amounts of  
425 carvacrol (>74%) (Salas et al., 2010) and another essential oil from the aerial parts of *T.*  
426 *capitata* collected in the south of Portugal showed the same trend with carvacrol  
427 accounting up to 75% (Palmeira-de-Oliveira et al., 2012). The potential toxicity of the *T.*  
428 *capitata* hydrolate towards *D. magna* was also assessed. Despite carvacrol being the

429 most abundant compound (98.1%), no toxic effects were observed up to 400 mg/L.  
430 Hydrolates are a by-product of the process of obtaining an essential oil (hydro- or steam-  
431 distillation) they usually contain lower concentrations of compounds than essential oils,  
432 and these are generally water-soluble compounds that end up as a residue of the  
433 process to obtain essential oils. This leads to usually softer scents and lower biological  
434 activity of these products when compared with the essential oils (Catty, 2001). The  
435 hydrolate was mainly composed of carvacrol. This composition is consistent with  
436 another study that compared the chemical composition of an essential oil and a  
437 hydrolate obtained from *T. capitata* that links this high abundance to the hydrophilic  
438 character of carvacrol (Moukhles et al., 2020). It has been shown that the yield of  
439 compounds present in an essential oil and the corresponding hydrolate from *T. capitata*  
440 is much lower in the hydrolate form (1.99% for the essential oil vs 0.45% for the  
441 hydrolate) (Moukhles et al., 2019). Although hydrolates are usually linked to lower  
442 bioactive activities, a recent study showed acute toxic effects of a hydrolate obtained  
443 from *Artemisia absinthium* towards *D. magna* at relatively low concentrations (EC<sub>50</sub>=  
444 0.24% of hydrolate dilution), and in this way, it is also important to evaluate potential  
445 toxic effects of these industry by-products to non-target organisms (Pino-Otín et al.,  
446 2019).

447

#### 448 **3.2.4 *Ocimum basilicum* hydrolate**

449 The *Ocimum basilicum* hydrolate also showed no acute toxic effects to *D. magna* in this  
450 study up to very high concentrations (8000 mg/L). The hydrolate composition revealed  
451 the presence of seven compounds. Eugenol (52.5%) and linalool (38.3%) were the major

452 compounds identified, but lower amounts of  $\alpha$ -pinene, eucalyptol (or 1,8-cineole),  
453 camphor,  $\alpha$ -terpineol and geraniol were also present. Although eugenol is reported to  
454 be acutely toxic to *D. magna* at very low concentrations, with one study reporting a 48h  
455 EC<sub>50</sub> of 0.70 mg/L (Gueretz et al., 2017), it has very low solubility in water (Baker and  
456 Grant, 2018) meaning it is not present at high concentrations in hydrolates. For linalool,  
457 an 48h EC<sub>50</sub> of 20 mg/L has been reported (Api et al., 2016), and in the ECHA database,  
458 a 48h EC<sub>50</sub> of 59 mg/L to *D. magna* is reported (ECHA). Similarly to eugenol, linalool is  
459 also poorly soluble in water, and in this way, the amount of the compound present in  
460 the hydrolate is not expected to be very high. These facts can explain of the lack of  
461 toxicity of the hydrolate from *O. basilicum* tested towards *D. magna* up to very high  
462 concentrations.

463

#### 464 **4. Conclusion**

465 Essential oils are important raw materials used in many industries for decades. Recently,  
466 hydrolates, which were commonly disregarded and categorized as a waste from the  
467 distillation process, have been increasingly considered, as they possess interesting  
468 bioactivities and add extra value under a circular economy approach. Despite being  
469 generally regarded as green and safe products, essential oils and other extracts obtained  
470 from plants may pose environmental risks to certain organisms. Considering the possible  
471 industrial applications of essential oils and hydrolates from *C. ladanifer*, *H. italicum*, *O.*  
472 *basilicum* and *T. capitata* and following the Precautionary Principle, the acute toxicity of  
473 these products was tested for their eco-safety using the model organism *D. magna*. All  
474 the hydrolates tested presented no risk to this organism. The essential oils from *H.*

475 *italicum* and *C. ladanifer* present low to no risk to *D. magna*. The essential oil from *T.*  
476 *capitata* showed moderate toxicity to *D. magna* and therefore precautions should be  
477 taken to avoid the contamination of water bodies when producing, handling, and using  
478 this product especially due to the high carvacrol content of the oil.

479

## 480 **Acknowledgments**

481 This work was supported by "INOVEP project – Innovation with Plant Extracts", I&DT  
482 projects for companies in collaboration with scientific entities, project number 33815,  
483 Centro2020. Further financial support was provided by Fundação Ciência e Tecnologia,  
484 IP to the Health Sciences Research Center, University of Beira Interior (CICS-UBI)  
485 through the project UID/Multi/00709/2019) and to the Comprehensive Health Research  
486 Centre (CHRC), University of Évora, through the project UIDP/04923/2020. C.A. Ferraz  
487 acknowledges the financial support from University of Beira Interior through the  
488 scholarship co-funded by the European Social Fund - P2020/POISE and the support by  
489 the Regional Direction for Higher Education from the Autonomous Region of Madeira  
490 and the Municipality of Câmara de Lobos.

491

## 492 **References**

- 493 Açıköz, M.A., 2020. Establishment of cell suspension cultures of *Ocimum basilicum* L. and  
494 enhanced production of pharmaceutical active ingredients. *Industrial Crops and Products* 148,  
495 112278, <https://doi.org/10.1016/j.indcrop.2020.112278>.
- 496 Aleksic Sabo, V., Knezevic, P., 2019. Antimicrobial activity of *Eucalyptus camaldulensis* Dehn.  
497 plant extracts and essential oils: A review. *Industrial Crops and Products* 132, 413-429,  
498 <https://doi.org/10.1016/j.indcrop.2019.02.051>.
- 499 Anastasiou, T.I., Mandalakis, M., Krigas, N., Vézignol, T., Lazari, D., Katharios, P., Dailianis, T.,  
500 Antonopoulou, E., 2020. Comparative evaluation of essential oils from medicinal-aromatic

501 plants of Greece: Chemical composition, antioxidant capacity and antimicrobial activity against  
502 bacterial fish pathogens. *Molecules* 25, 10.3390/molecules25010148.

503 Antunes Viegas, D., Palmeira-de-Oliveira, A., Salgueiro, L., Martinez-de-Oliveira, J., Palmeira-  
504 de-Oliveira, R., 2014. *Helichrysum italicum*: from traditional use to scientific data. *Journal of*  
505 *ethnopharmacology* 151, 54-65, 10.1016/j.jep.2013.11.005.

506 Api, A.M., Belsito, D., Bhatia, S., Bruze, M., Calow, P., Dagli, M.L., Dekant, W., Fryer, A.D.,  
507 Kromidas, L., La Cava, S., Lalko, J.F., Lapczynski, A., Liebler, D.C., Miyachi, Y., Politano, V.T.,  
508 Ritacco, G., Salvito, D., Schultz, T.W., Shen, J., Sipes, I.G., Wall, B., Wilcox, D.K., 2016. RIFM  
509 fragrance ingredient safety assessment, l-linalool, CAS Registry Number 126-91-0. *Food and*  
510 *Chemical Toxicology* 97, S11-S24, <https://doi.org/10.1016/j.fct.2015.12.014>.

511 ASTM, 1997. ASTM E1193-97, Standard Guide for Conducting Daphnia magna Life-Cycle  
512 Toxicity Tests. ASTM International, [www.astm.org](http://www.astm.org).

513 Baker, B.P., Grant, J.A., 2018. Eugenol profile.

514 Barrajón-Catalán, E., Tomás-Menor, L., Morales-Soto, A., Bruñá, N.M., López, D.S., Segura-  
515 Carretero, A., Micol, V., 2016. Rockroses (*Cistus* sp.) oils, *Essential Oils in Food Preservation,*  
516 *Flavor and Safety*, pp. 649-658.

517 Catty, S., 2001. *Hydrosols: the next aromatherapy*. Inner Traditions/Bear & Co.

518 Charfi, S., Boujida, N., Abrini, J., Senhaji, N.S., 2019. Study of chemical composition and  
519 antibacterial activity of Moroccan *Thymbra capitata* essential oil and its possible use in orange  
520 juice conservation. *Materials Today: Proceedings* 13, 706-712,  
521 <https://doi.org/10.1016/j.matpr.2019.04.031>.

522 Collin, G., Gagnon, H., 2016. Chemical composition and stability of the hydrosol obtained  
523 during the production of essential oils. III. The case of *Myrica gale* L., *Comptonia peregrina* (L.)  
524 Coulter and *Ledum groenlandicum* Retzius. *American Journal of Essential Oils and Natural*  
525 *Products* 4, 07-19.

526 Costa, P., Loureiro, J.M., Teixeira, M.A., Rodrigues, A.E., 2015. Extraction of aromatic volatiles  
527 by hydrodistillation and supercritical fluid extraction with CO<sub>2</sub> from *Helichrysum italicum*  
528 subsp. *picardii* growing in Portugal. *Industrial Crops and Products* 77, 680-683,  
529 <https://doi.org/10.1016/j.indcrop.2015.09.042>.

530 Costa, R., De Fina, M., Valentino, M., Crupi, M., Mondello, L., 2007. Application of a new GC-  
531 MS library, designed with a retention index filter tool, to the analysis of the essential oil of  
532 *Cistus ladanifer*, I International Medicinal and Aromatic Plants Conference on Culinary Herbs  
533 826, pp. 271-276.

534 Djihane, B., Wafa, N., Elkhamssa, S., Maria, A.E., Mihoub, Z.M., 2017. Chemical constituents of  
535 *Helichrysum italicum* (Roth) G. Don essential oil and their antimicrobial activity against Gram-  
536 positive and Gram-negative bacteria, filamentous fungi and *Candida albicans*. *Saudi*  
537 *Pharmaceutical Journal* 25, 780-787.

538 Dzamic, A.M., Mileski, K.S., Ciric, A.D., Ristic, M.S., Sokovic, M.D., Marin, P.D., 2019. Essential  
539 Oil Composition, Antioxidant and Antimicrobial Properties of Essential Oil and Deodorized  
540 Extracts of *Helichrysum italicum* (Roth) G. Don. *Journal of Essential Oil Bearing Plants* 22, 493-  
541 503.

542 Ebert, D., 2005. Introduction to the Ecology, Epidemiology, and Evolution of Parasitism in  
543 *Daphnia*.

544 ECHA, Carvacrol. Registered-dossier/23562/6/2/6.

545 ECHA, linalool; 3,7-dimethyl-1,6-octadien-3-ol; dl-linalool. Registered-dossier/14501/6/2/4.

546 ECHA, 2016. Essential oil of *Cistus ladaniferus* L (Cistaceae) obtained from stems and leaves by  
547 distillation. Registered-dossier/21887/6/2/4.

548 Europe, C.o., 1997. European pharmacopoeia. Council of Europe, Strasbourg.

549 Frazão, D.F., Raimundo, J.R., Domingues, J.L., Quintela-Sabaris, C., Gonçalves, J.C., Delgado, F.,  
550 2018. *Cistus ladanifer* (Cistaceae): a natural resource in Mediterranean-type ecosystems.  
551 *Planta* 247, 289-300, 10.1007/s00425-017-2825-2.

552 Gomes, P.B., Mata, V.G., Rodrigues, A., 2005. Characterization of the Portuguese-grown *Cistus*  
553 *ladanifer* essential oil. *Journal of Essential Oil Research* 17, 160-165.

554 Granato, D., Nunes, D.S., Barba, F.J., 2017. An integrated strategy between food chemistry,  
555 biology, nutrition, pharmacology, and statistics in the development of functional foods: A  
556 proposal. *Trends in Food Science & Technology* 62, 13-22,  
557 <https://doi.org/10.1016/j.tifs.2016.12.010>.

558 Greche, H., Mrabet, N., Zrira, S., Ismaili-Alaoui, M., Benjilali, B., Boukir, A., 2009. The volatiles  
559 of the leaf oil of *Cistus ladanifer* L. var. *albiflorus* and labdanum extracts of moroccan origin  
560 and their antimicrobial activities. *Journal of Essential Oil Research* 21, 166-173.

561 Gueretz, J.S., Somensi, C.A., Martins, M.L., Souza, A.P.d., 2017. Evaluation of eugenol toxicity in  
562 bioassays with test-organisms. *Ciência Rural* 47.

563 Gülz, P.-G., Kobold, U., Michaelis, K., Vostrowsky, O., 1984. The composition of terpene  
564 hydrocarbons in the essential oils from leaves of four *Cistus* species. *Zeitschrift für*  
565 *Naturforschung A* 39, 699-704.

566 Hamdi, A., Majouli, K., Vander Heyden, Y., Flamini, G., Marzouk, Z., 2017. Phytotoxic activities  
567 of essential oils and hydrosols of *Haplophyllum tuberculatum*. *Industrial Crops and Products*  
568 97, 440-447, <https://doi.org/10.1016/j.indcrop.2016.12.053>.

569 Harhaun, R., Kunik, O., Saribekova, D., Lazzara, G., 2020. Biologically active properties of plant  
570 extracts in cosmetic emulsions. *Microchemical Journal* 154, 104543,  
571 <https://doi.org/10.1016/j.microc.2019.104543>.

572 **ISO, 2012. ISO 6341:2012. Water quality—Determination of the inhibition of the mobility of**  
573 ***Daphnia magna* Straus (Cladocera, Crustacea)—Acute toxicity test.**

574 ISO, 2013. ISO 9235:2013 Aromatic Natural Raw Materials - Vocabulary.

575 Kladar, N.V., Anačkov, G.T., Rat, M.M., Srd Strok Signenovic, B.U., Grujic, N.N., Šefer, E.I., Božin,  
576 B.N., 2015. Biochemical characterization of *helichrysum italicum* (Roth) G.Don subsp. *italicum*  
577 (Asteraceae) from montenegro: Phytochemical screening, chemotaxonomy, and antioxidant  
578 properties. *Chemistry and Biodiversity* 12, 419-431, 10.1002/cbdv.201400174.

579 Mariotti, J., Tomi, F., Casanova, J., Costa, J., Bernardini, A., 1997. Composition of the essential  
580 oil of *Cistus ladaniferus* L. cultivated in Corsica (France). *Flavour and fragrance journal* 12, 147-  
581 151.

582 Marwat, S.K., Fazal Ur, R., Khan, M.S., Ghulam, S., Anwar, N., Mustafa, G., Usman, K., 2011.  
583 Phytochemical constituents and pharmacological activities of sweet Basil-*Ocimum basilicum* L.  
584 (Lamiaceae). *Asian Journal of Chemistry* 23, 3773-3782.

585 Moukhles, A., Belcadi, H., Raissouni, I., Ben driss, A., Mansour, A.I., 2020. Chemical  
586 Composition, in vitro Antibacterial Activity and Corrosion Inhibition of Essential Oil and  
587 Hydrolat Extract from Aerial Parts of *Thymbra capitata* (L.) Cav Harvested at Northern  
588 Morocco. *Journal of Essential Oil Bearing Plants* 23, 375-389.

589 Moukhles, A., Charfi, S., Zantar, S., Toukour, L., Mansour, A.I., 2019. Seasonal variation in yield  
590 and chemical composition of Moroccan *Thymbra capitata* (L.) Cav. essential oil and its  
591 corresponding hydrolat extracted essential oil. *Moroccan Journal of Chemistry* 7, 7-2 (2019)  
592 2346-2353.

593 Neves, A., Marto, J., Duarte, A., Gonçalves, L.M., Pinto, P., Figueiredo, A.C., Ribeiro, H.M., 2017.  
594 Characterization of Portuguese *Thymbra capitata*, *Thymus caespitius* and *Myrtus communis*  
595 essential oils in topical formulations. *Flavour and Fragrance Journal* 32, 392-402,  
596 10.1002/ffj.3393.

597 OECD, 2004. Test No. 202: *Daphnia* sp. Acute Immobilisation Test.

598 OECD, 2019. Guidance Document on Aquatic Toxicity Testing of Difficult Substances and  
599 Mixtures.

600 Oliva, A., Garzoli, S., Sabatino, M., Tadić, V., Costantini, S., Ragno, R., Božović, M., 2020.  
601 Chemical composition and antimicrobial activity of essential oil of *Helichrysum italicum* (Roth)  
602 G. Don fil. (Asteraceae) from Montenegro. *Natural Product Research* 34, 445-448,  
603 10.1080/14786419.2018.1538218.

604 Palmeira-de-Oliveira, A., Gaspar, C., Palmeira-de-Oliveira, R., Silva-Dias, A., Salgueiro, L.,  
605 Cavaleiro, C., Pina-Vaz, C., Martinez-de-Oliveira, J., Queiroz, J.A., Rodrigues, A.G., 2012. The  
606 anti-*Candida* activity of *Thymbra capitata* essential oil: effect upon pre-formed biofilm. *J*  
607 *Ethnopharmacol* 140, 379-383, 10.1016/j.jep.2012.01.029.

608 Papaefthimiou, D., Papanikolaou, A., Falara, V., Givanoudi, S., Kostas, S., Kanellis, A.K., 2014.  
609 Genus *Cistus*: a model for exploring labdane-type diterpenes' biosynthesis and a natural source  
610 of high value products with biological, aromatic, and pharmacological properties. *Frontiers in*  
611 *Chemistry* 2, 10.3389/fchem.2014.00035.

612 Pino-Otín, M.R., Ballesteros, D., Navarro, E., González-Coloma, A., Val, J., Mainar, A.M., 2019.  
613 Ecotoxicity of a novel biopesticide from *Artemisia absinthium* on non-target aquatic organisms.  
614 *Chemosphere* 216, 131-146, 10.1016/j.chemosphere.2018.09.071.

615 Qaralleh, H., 2019. Thymol rich *thymbra capitata* essential oil inhibits quorum sensing,  
616 virulence and biofilm formation of beta lactamase producing *Pseudomonas aeruginosa*.  
617 *Natural Product Sciences* 25, 172-180, 10.20307/nps.2019.25.2.172.

618 Rafinska, K., Pomastowski, P., Rudnicka, J., Krakowska, A., Maruska, A., Narkute, M., Buszewski,  
619 B., 2019. Effect of solvent and extraction technique on composition and biological activity of  
620 *Lepidium sativum* extracts. *Food chemistry* 289, 16-25, 10.1016/j.foodchem.2019.03.025.

621 Ragab, G.A., Saad-Allah, K.M., 2020. Green synthesis of sulfur nanoparticles using *Ocimum*  
622 *basilicum* leaves and its prospective effect on manganese-stressed *Helianthus annuus* (L.)  
623 seedlings. *Ecotoxicology and environmental safety* 191, 10.1016/j.ecoenv.2020.110242.

624 Raimundo, J.R., Frazao, D.F., Domingues, J.L., Quintela-Sabaris, C., Dentinho, T.P., Anjos, O.,  
625 Alves, M., Delgado, F., 2018. Neglected Mediterranean plant species are valuable resources:  
626 the example of *Cistus ladanifer*. *Planta* 248, 1351-1364, 10.1007/s00425-018-2997-4.

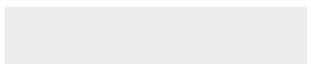
627 Renu, S., Shivashangari, K.S., Ravikumar, V., 2019. Incorporated plant extract fabricated  
628 silver/poly-D,L-lactide-co-glycolide nanocomposites for antimicrobial based wound healing.  
629 *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 117673,  
630 <https://doi.org/10.1016/j.saa.2019.117673>.

631 Riani, L., Macedo, A., Chedier, L., Pimenta, D., 2017. Chemical analysis of essential oil and  
632 hydrolates of leaves, inflorescences and stems of *piper chimonanthifolium* kunth. *Rev. Virtual*  
633 *Quim* 9, 1560-1569.

- 634 Robles, C., Bousquet-Mélou, A., Garzino, S., Bonin, G., 2003. Comparison of essential oil  
635 composition of two varieties of *Cistus ladanifer*. *Biochemical Systematics and Ecology* 31, 339-  
636 343.
- 637 Rossi, P.-G., Berti, L., Panighi, J., Luciani, A., Maury, J., Muselli, A., Serra, D.d.R., Gonny, M.,  
638 Bolla, J.-M., 2007. Antibacterial action of essential oils from Corsica. *Journal of Essential Oil*  
639 *Research* 19, 176-182.
- 640 Salas, J.B., Téllez, T.R., Alonso, M.J.P., Pardo, F.M.V., de los Ángeles Cases Capdevila, M.,  
641 Rodríguez, C.G., 2010. Chemical composition and antioxidant activity of the essential oil of  
642 *Thymbra capitata* (L.) Cav. in Spain. *Acta botanica gallica* 157, 55-63.
- 643 Sarkic, A., Stappen, I., 2018. Essential Oils and Their Single Compounds in Cosmetics—A Critical  
644 Review. *Cosmetics* 5, 11.
- 645 Tavares, C.S., Martins, A., Faleiro, M.L., Miguel, M.G., Duarte, L.C., Gameiro, J.A., Roseiro, L.B.,  
646 Figueiredo, A.C., 2020. Bioproducts from forest biomass: Essential oils and hydrolates from  
647 wastes of *Cupressus lusitanica* Mill. and *Cistus ladanifer* L. *Industrial Crops and Products* 144,  
648 112034, <https://doi.org/10.1016/j.indcrop.2019.112034>.
- 649 Verdeguer, M., Blázquez, M.A., Boira, H., 2012. Chemical composition and herbicidal activity of  
650 the essential oil from a *Cistus ladanifer* L. population from Spain. *Natural product research* 26,  
651 1602-1609.
- 652 Viuda-Martos, M., Sendra, E., Pérez-Alvarez, J.A., Fernández-López, J., Amensour, M., Abrini, J.,  
653 2011. Identification of flavonoid content and chemical composition of the essential oils of  
654 Moroccan herbs: myrtle (*Myrtus communis* L.), rockrose (*Cistus ladanifer* L.) and Montpellier  
655 cistus (*Cistus monspeliensis* L.). *Journal of Essential Oil Research* 23, 1-9.
- 656 Zidane, H., Elmiz, M., Aouinti, F., Tahani, A., Wathelet, J., Sindic, M., Elbachiri, A., 2013.  
657 Chemical composition and antioxidant activity of essential oil, various organic extracts of *Cistus*  
658 *ladanifer* and *Cistus libanotis* growing in Eastern Morocco. *African Journal of Biotechnology* 12,  
659 5314-5320.



Click here to access/download  
**e-component**  
Graphical abstract.emf



## **Credit Author Statement**

**Celso Afonso Ferraz:** Investigation, Visualization, Writing - Original Draft. **Ana Catarina**

**Sousa:** Conceptualization, Methodology, Supervision, Writing - Review & Editing.

**Débora Caramelo:** Investigation. **Fernanda Delgado:** Resources, Supervision, Writing -

Review & Editing. **Ana Palmeira de Oliveira:** Funding acquisition, Writing - Review &

Editing. **M. Ramiro Pastorinho:** Conceptualization, Resources, Supervision, Writing -

Review & Editing.

**Declaration of interests**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

The authors declare the following financial interests/personal relationships which may be considered as potential competing interests: