

RESEARCH ARTICLE

**Insect-based chitin and chitosan from whole body sources and rearing by-products: extraction, physicochemical, structural and bioactivity characterisation**

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**Supplementary material**

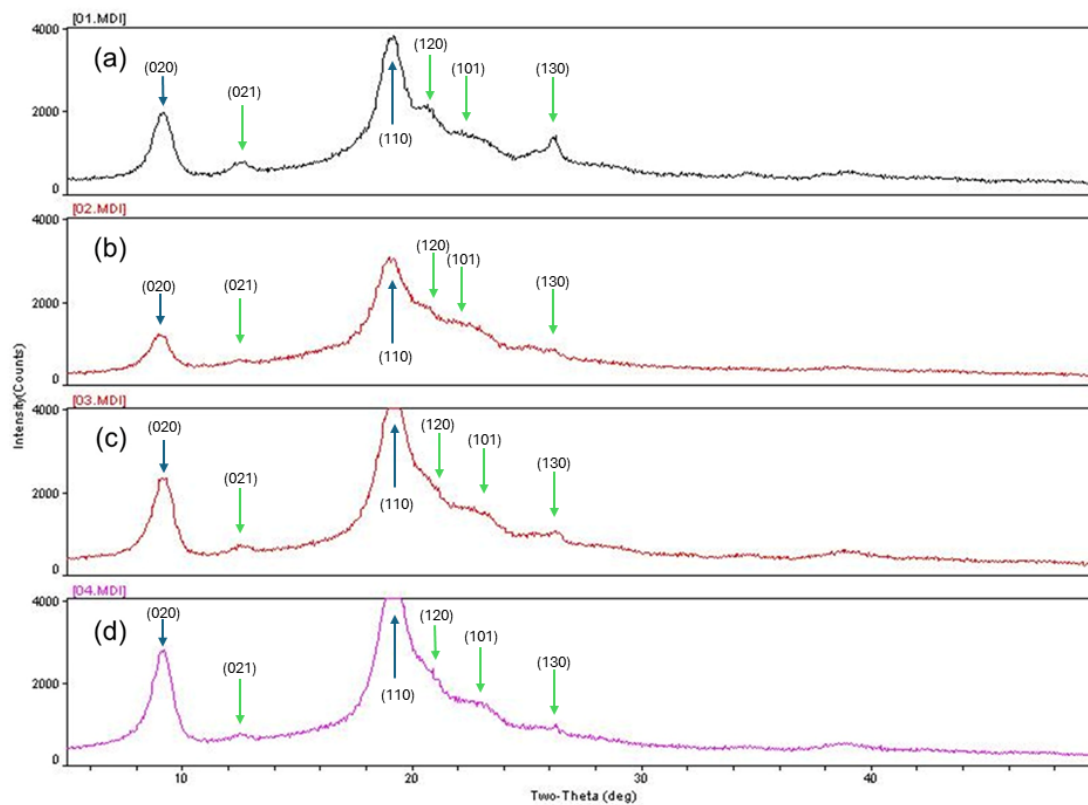


FIGURE S1 XRD patterns of chitin from various sources: (a) commercial chitin, (b) *T. molitor* larvae (TM), (c) *A. domesticus* (AD) and (d) *A. domesticus* legs & wings (house cricket L&W); Blue arrows indicate major peaks on the plane and green arrows indicate minor peaks on the plane.

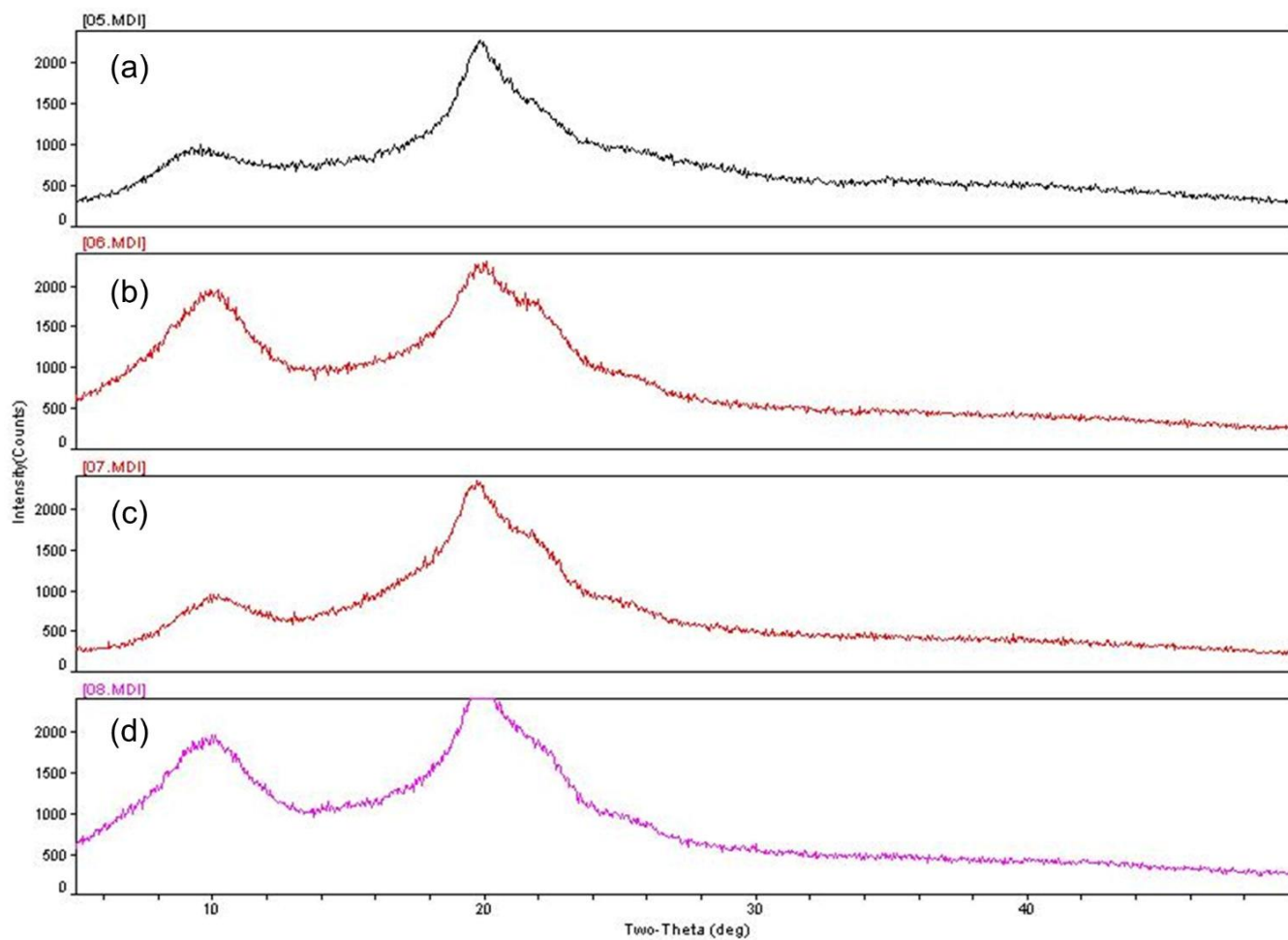


FIGURE S2 XRD patterns of chitosan from various sources: (a) commercial chitin, (b) *T. molitor* larvae (TM), (c) *A. domesticus* (AD) and (d) *A. domesticus* legs & wings (house cricket L&W).

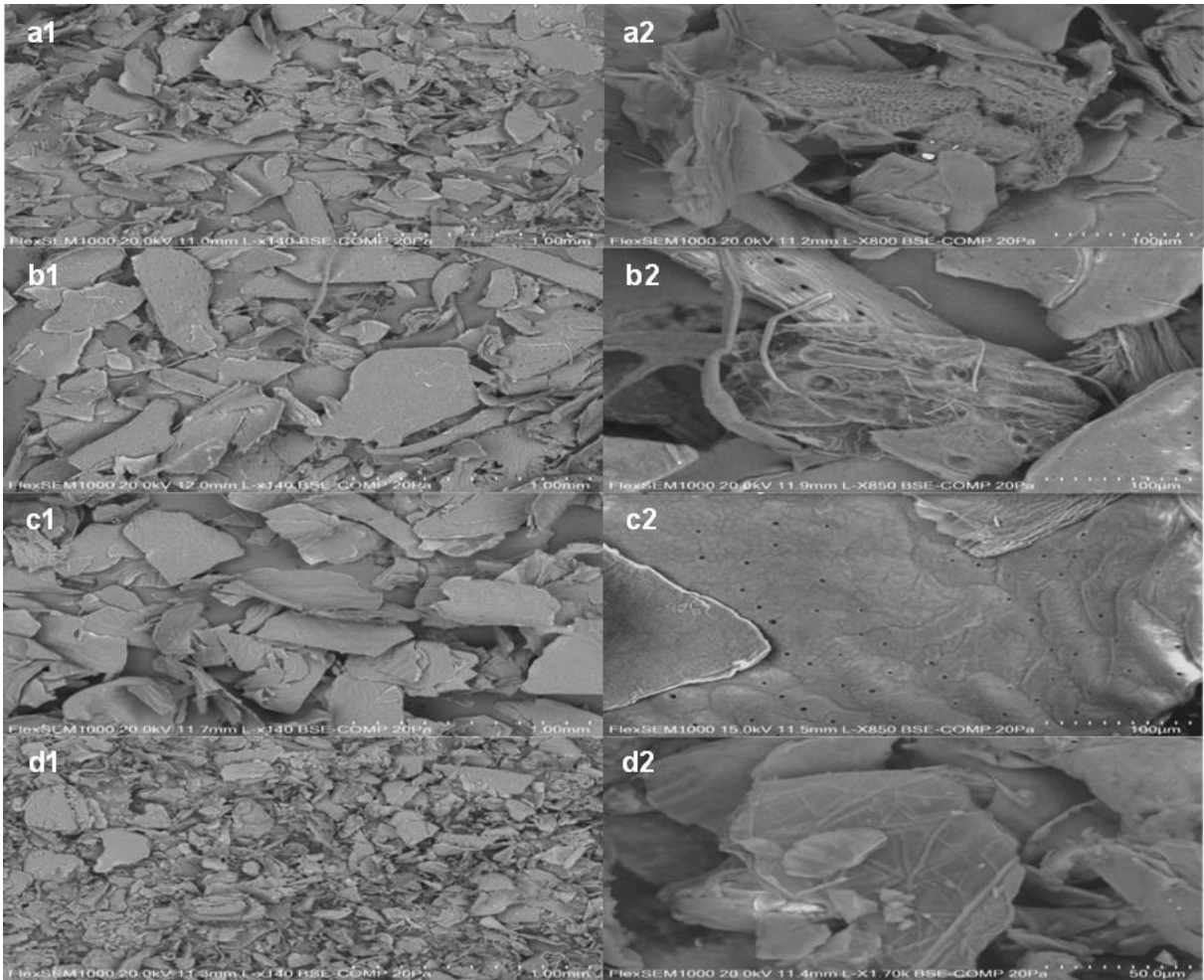


FIGURE S3 Surface morphology of chitin samples (a) *A. domesticus* (AD), (b) *A. domesticus* legs & wings (house cricket L&W), (c) *T. molitor* larvae (TM), (d) commercial chitin from SEM images at different magnifications (1 = x170; 2 = x1.7k).

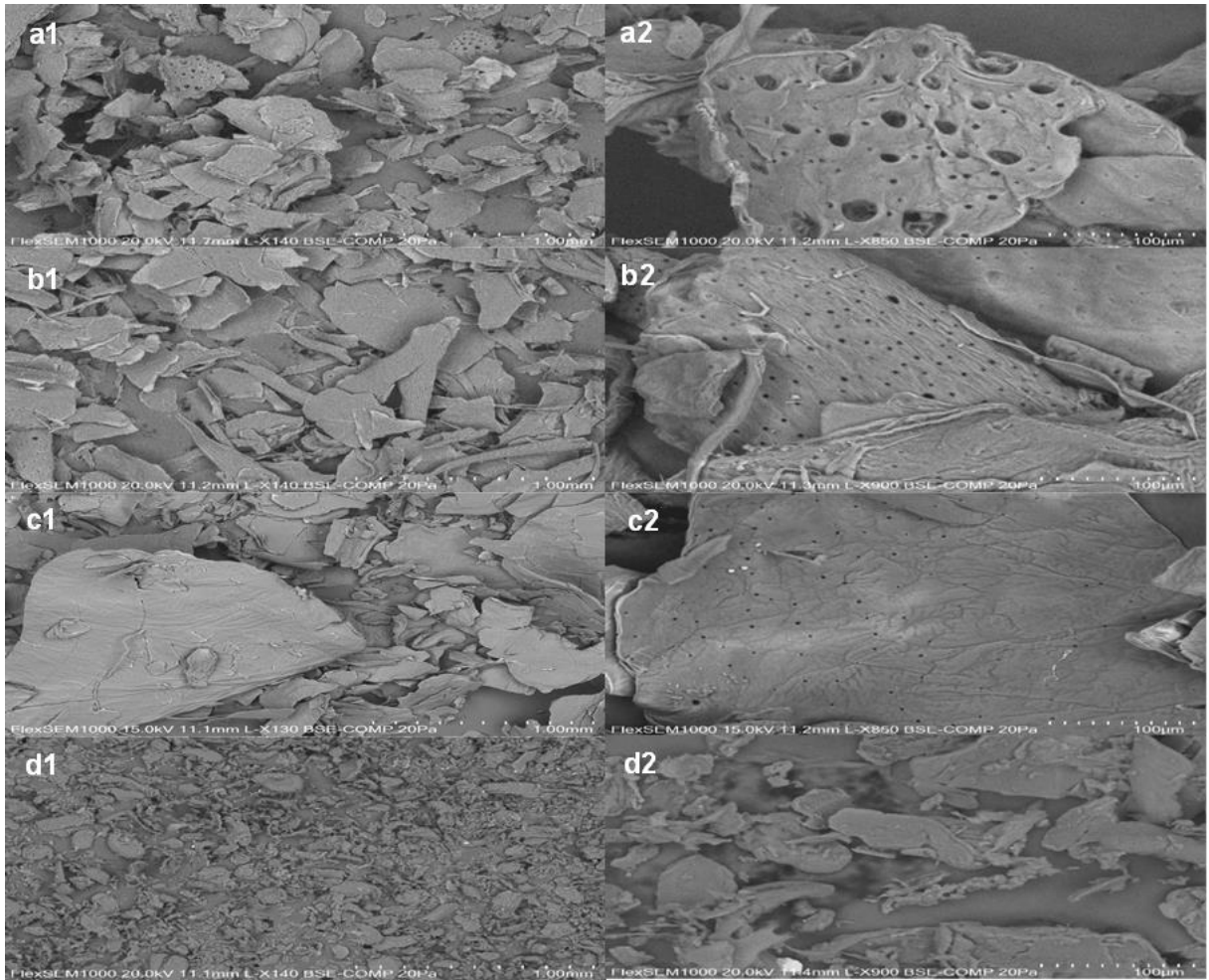


FIGURE S4 Surface morphology of chitosan (a) *A. domesticus* (AD), (b) *A. domesticus* legs & wings (house cricket L&W), (c) *T. molitor* larvae (TM), (d) – commercial chitin from SEM images at different magnifications (1 = x170; 2 = x1.7k).

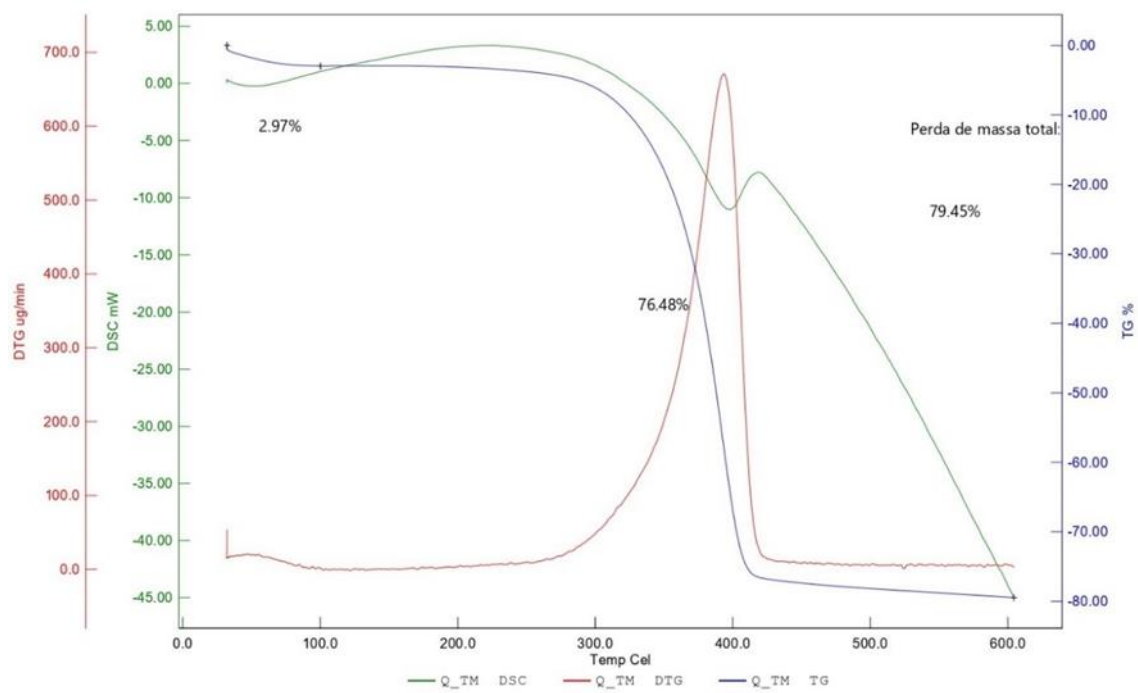


FIGURE S5 DTG, TG and DSC curves of *T. molitor* larvae (TM) chitin.

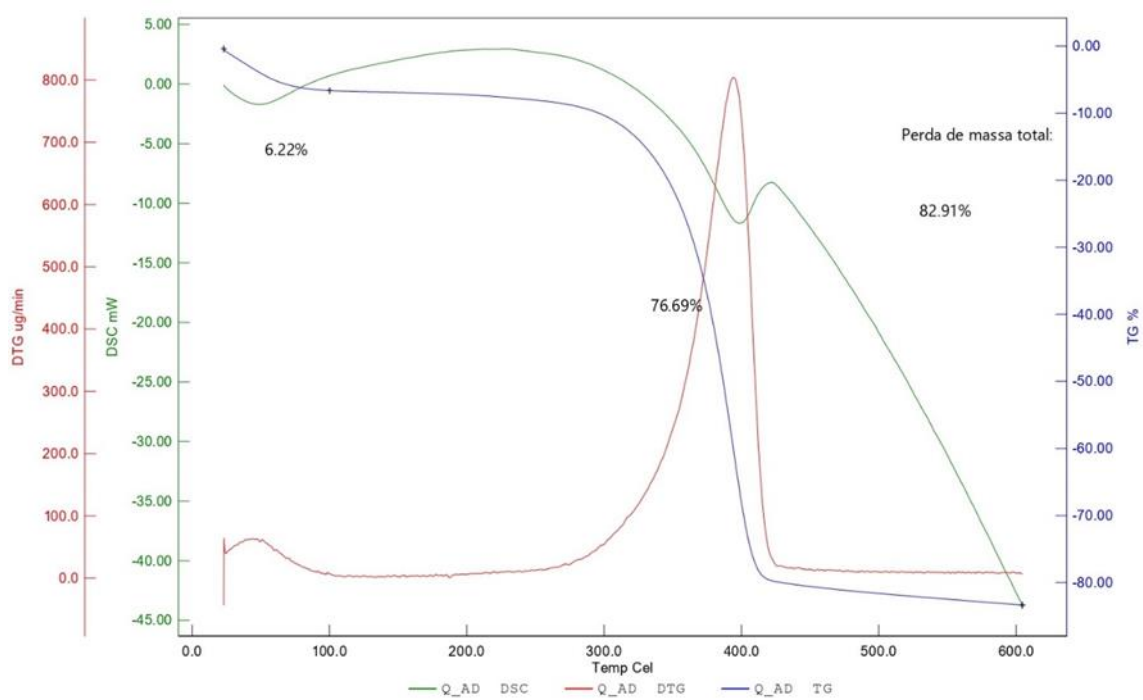


FIGURE S6 DTG, TG and DSC curves of *A. domesticus* (AD) chitin.

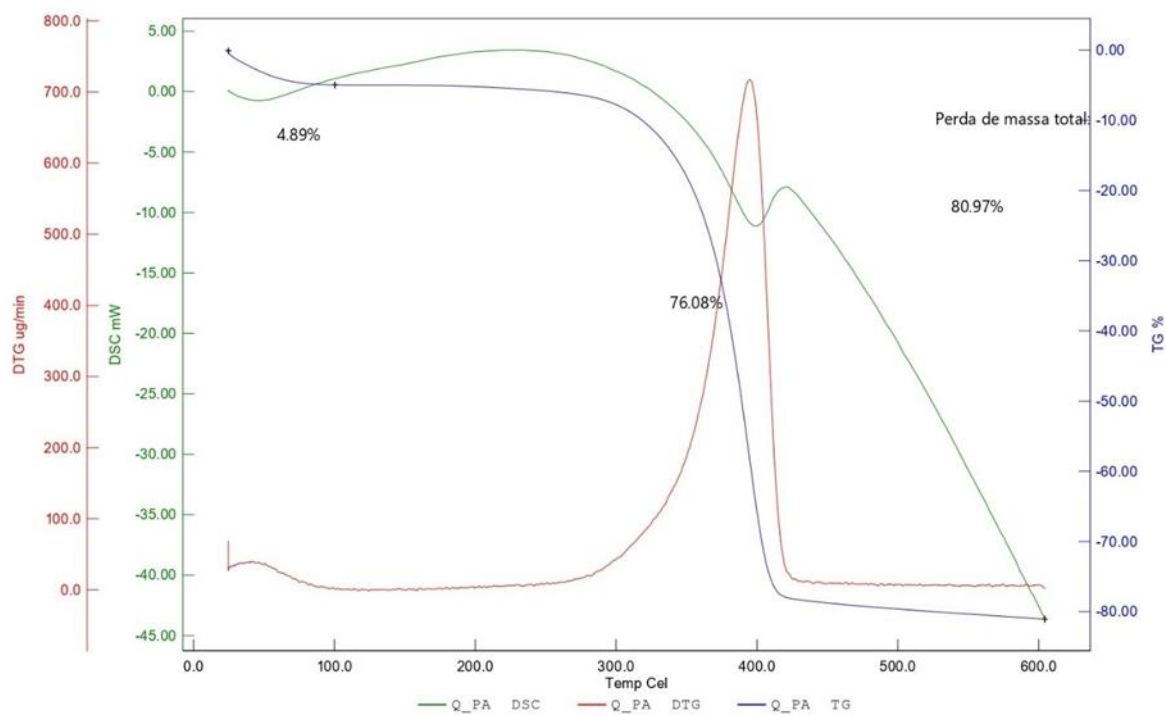


FIGURE S7 DTG, TG and DSC curves of *A. domesticus* legs & wings (house cricket L&W) chitin.

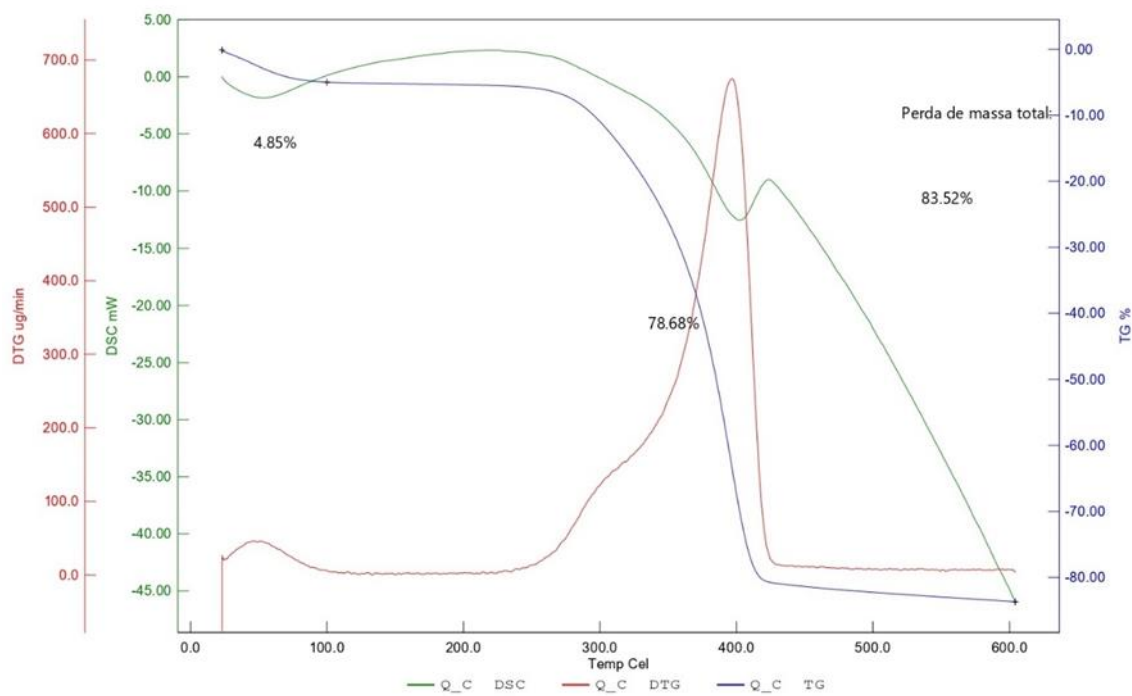


FIGURE S8 DTG, TG and DSC curves of commercial chitin.

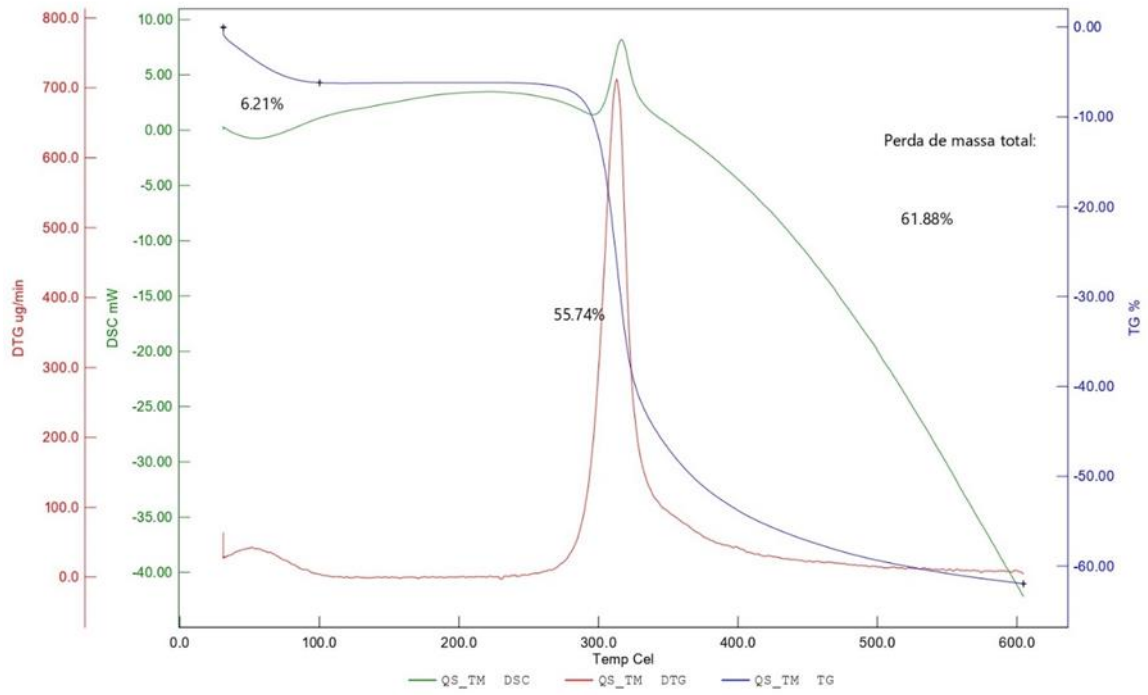


FIGURE S9 DTG, TG and DSC curves of *T. molitor* larvae (TM) chitosan.

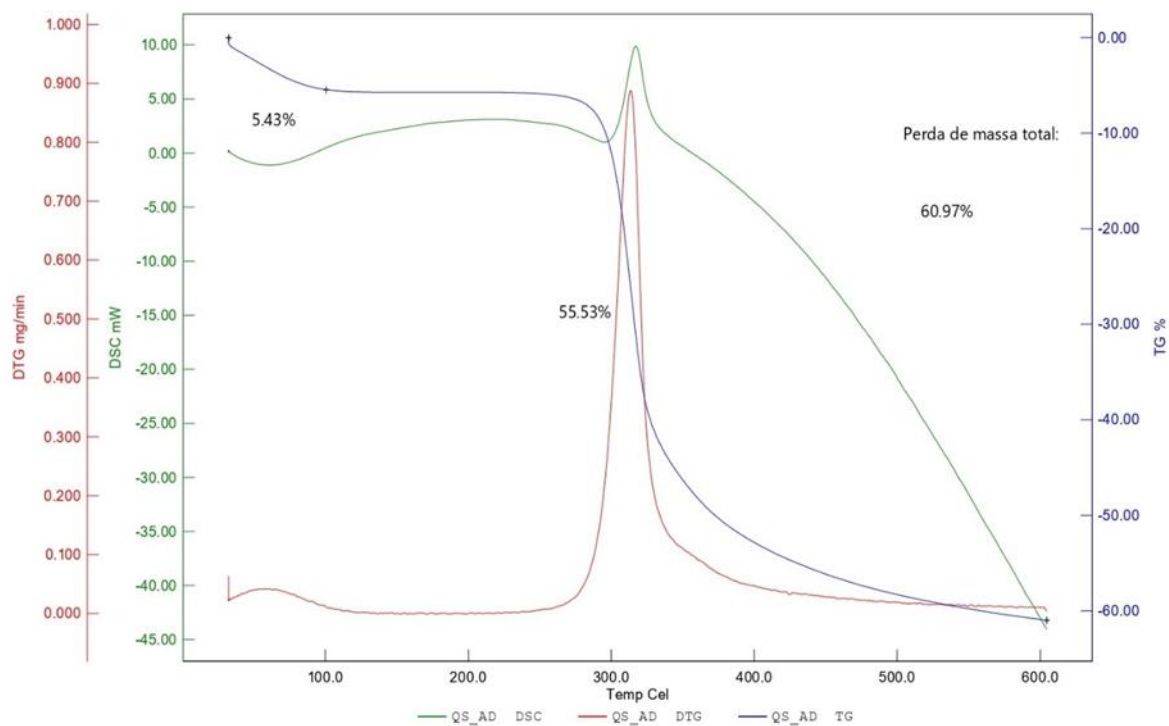


FIGURE S10 DTG, TG and DSC curves of *A. domesticus* (AD) chitosan.

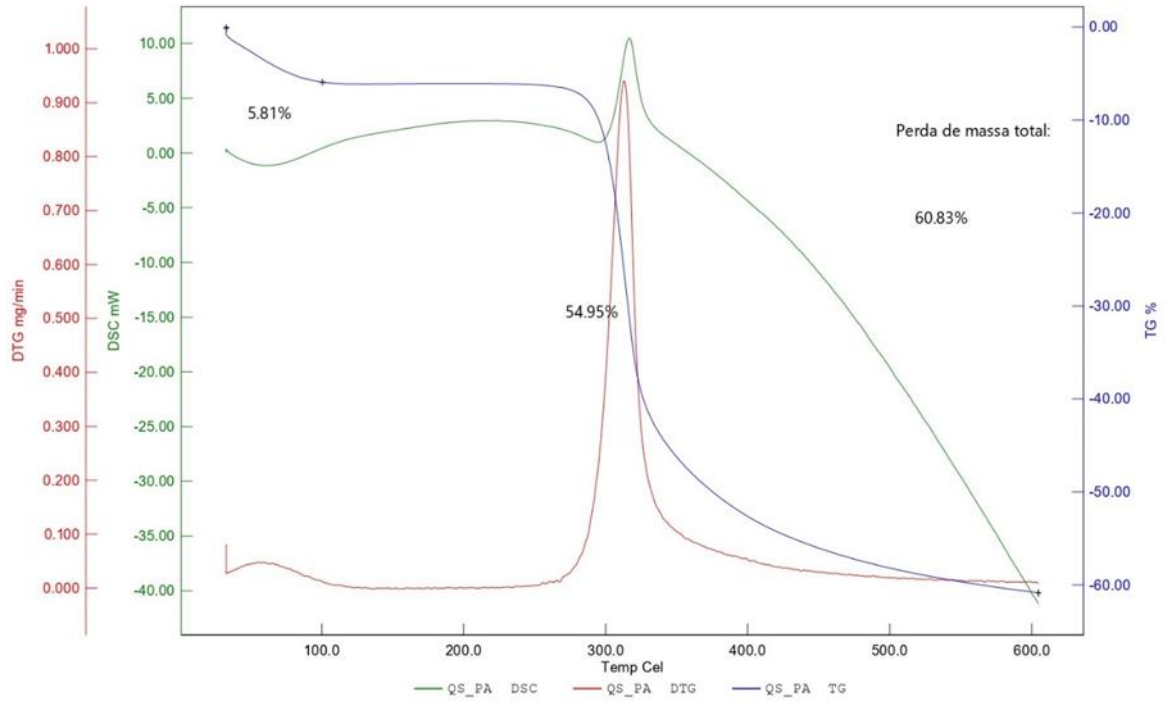


FIGURE S11 DTG, TG and DSC curves of *A. domesticus* legs & wings (house cricket L&W) chitosan.

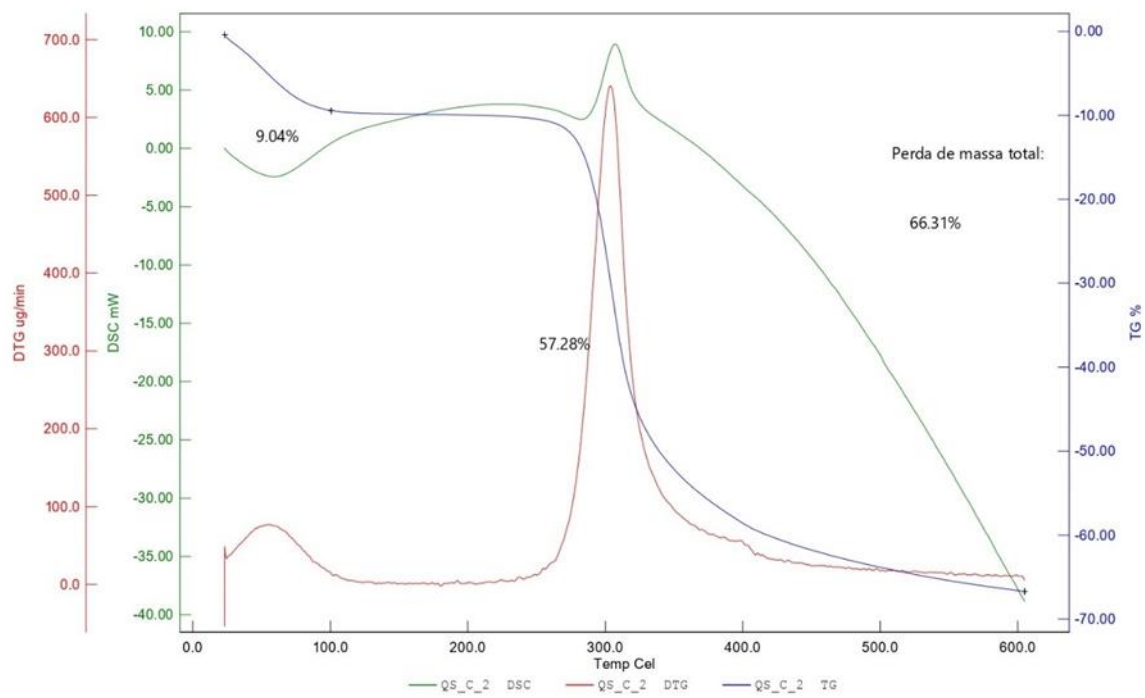


FIGURE S12 DTG, TG and DSC curves of commercial chitosan.

TABLE S1 Color (CIE-Lab\*)coordinates and whiteness index (n = 6, mean  $\pm$  SD) of chitosan samples

Samples	Chitosan			
	$L^*$	$a^*$	$b^*$	WI
<i>T. molitor</i> larvae (TM)	77.61 $\pm$ 1.19 <sup>b</sup>	2.93 $\pm$ 0.07 <sup>b</sup>	13.95 $\pm$ 0.10 <sup>d</sup>	73.45 $\pm$ 1.04 <sup>b</sup>
<i>A. domesticus</i> (AD)	71.20 $\pm$ 0.17 <sup>d</sup>	3.50 $\pm$ 0.8 <sup>a</sup>	15.43 $\pm$ 0.15 <sup>c</sup>	67.13 $\pm$ 0.20 <sup>d</sup>
<i>A. domesticus</i> by- products (house cricket L&W)	75.03 $\pm$ 0.24 <sup>c</sup>	3.13 $\pm$ 0.15 <sup>b</sup>	16.64 $\pm$ 0.65 <sup>b</sup>	69.83 $\pm$ 0.24 <sup>c</sup>
Commercial	101.07 $\pm$ 2.37 <sup>a</sup>	1.87 $\pm$ 0.21 <sup>c</sup>	17.22 $\pm$ 0.22 <sup>a</sup>	82.51 $\pm$ 0.24 <sup>a</sup>

<sup>a, b, c, d</sup>Homogenous groups in each column according to Tukey's *post hoc* test ( $P < 0.05$ ).

TABLE S2 Wavenumber of the different peaks attributed to functional groups characteristics of chitin

Functional group and vibration modes	Classification	<i>T. molitor</i> larvae (TM)	<i>A. domesticus</i> (AD)	<i>A. domesticus</i> by-products (L&W)	Commercial
O-H stretching	–	3,437	3,436	3,443	3,359
N-H stretching	–	3,097; 3,254	3,092; 3,255	3,097; 3,253	3,087; 3,254
CH <sub>3</sub> symmetric stretch and CH <sub>2</sub> asymmetric stretch	Aliphatic compounds	2,930	2,957	2,926	2,956
CH <sub>3</sub> symmetric stretch	Aliphatic compound	2,875	2,882	2,871	2,879
C=O secondary amide stretch	Amide I	1,651	1,651	1,652	1,652
C=O secondary amide stretch	Amide I	1,620	1,620	1,620	1,622
N-H bend, C-N stretch	Amide II	1,550	1,550	1,550	1,551
CH <sub>2</sub> ending and CH <sub>3</sub> deformation	–	1,422	1,422	1,424	1,421
CH bend, CH <sub>3</sub> symmetric deformation	–	1,372	1,373	1,372	1,373

CH <sub>2</sub> wagging	Amide III, components of protein	1,306	1,307	1,305	1,308
Asymmetric bridge oxygen stretching		1,154	1,153	1,155	1,153
Asymmetric in- phase ring stretching mode	Saccharide rings	1,114	1,112	1,115	1,111
C-O-C asymmetric stretch in phase ring		1,067	1,066	1,067	1,066
C-O asymmetric stretch in phase ring	–	1,008	1,012	1,006	1,013
CH <sub>3</sub> wagging	Along chain	949	950	949	950
CH ring stretching	Saccharide rings	894	895	894	894

TABLE S3 Lipid peroxidation inhibition (% , mean  $\pm$  SD) for chitosan samples and control (BHT) according to  $\beta$ -carotene bleaching test

Samples	Lipid peroxidation inhibition (%)					
	50 mg/L	100 mg/L	250 mg/L	500 mg/L	750 mg/L	1000 mg/L
<i>T. molitor</i> larvae (TM)	2.5 $\pm$ 1.2 <sup>b</sup>	14.5 $\pm$ 0.3 <sup>b</sup>	29.4 $\pm$ 0.5 <sup>c</sup>	36.3 $\pm$ 0.8 <sup>b</sup>	36.3 $\pm$ 0.8 <sup>b</sup>	42.1 $\pm$ 0.3 <sup>c</sup>
<i>A. domesticus</i> (AD)	4.5 $\pm$ 2.8 <sup>b</sup>	17.9 $\pm$ 0.9 <sup>b</sup>	34.4 $\pm$ 1.8 <sup>b</sup>	43.2 $\pm$ 1.8 <sup>ab</sup>	50.3 $\pm$ 0.6 <sup>b</sup>	54.5 $\pm$ 0.9 <sup>ab</sup>
<i>A. domesticus</i> by-products (L&W)	2.8 $\pm$ 1.7 <sup>b</sup>	16.9 $\pm$ 2.7 <sup>b</sup>	31.0 $\pm$ 1.4 <sup>bc</sup>	33.1 $\pm$ 1.4 <sup>b</sup>	39.5 $\pm$ 1.4 <sup>a</sup>	46.1 $\pm$ 5.5 <sup>bc</sup>
Commercial	15.5 $\pm$ 1.4 <sup>a</sup>	26.4 $\pm$ 0.4 <sup>a</sup>	44.9 $\pm$ 0.2 <sup>a</sup>	52.8 $\pm$ 0.4 <sup>a</sup>	55.2 $\pm$ 1.9 <sup>a</sup>	62.4 $\pm$ 0.1 <sup>a</sup>
BHT	43.4 $\pm$ 0.6	53.8 $\pm$ 0.7	71.1 $\pm$ 0.8	82.1 $\pm$ 1.1	90.3 $\pm$ 1.6	93.4 $\pm$ 3.8

<sup>a, b, c</sup>Homogenous groups in each column according to Tukey's *post hoc* test ( $P < 0.05$ ).