

# Optimizing the choice of microsatellites for fingerprinting *Eucalyptus*

L. SANCHEZ<sup>1</sup>, M.M. RIBEIRO<sup>2</sup>, C. RIBEIRO<sup>3</sup>, J.A. ARAÚJO<sup>3</sup>, N. BORRALHO<sup>3</sup>, C.M. MARQUES<sup>3</sup>

<sup>1</sup> *Unité Amélioration, Génétique et Physiologie Forestières, INRA Centre d'Orléans, 45166 Olivet, France*

<sup>2</sup> *Unidade Departamental de Silvicultura e Recursos Naturais, Escola Superior Agrária, 6001-909 Castelo Branco, Portugal*

<sup>3</sup> *RAIZ-Centro de Investigação Florestal, ITQB II, 2781-901 Oeiras, Portugal*

E-mail: leopoldo.sanchez@orleans.inra.fr



## PURPOSE OF THE STUDY

### [A] What is the Minimum set of Discriminant Markers (MSDM) ...

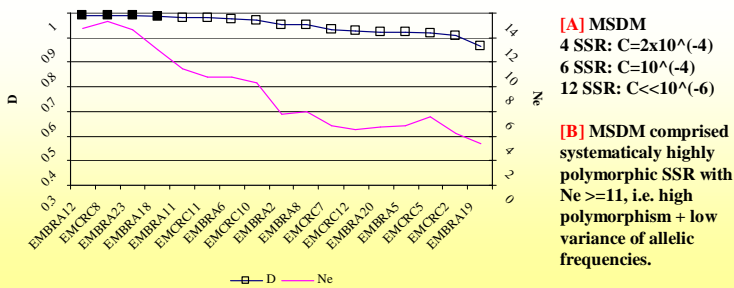
- ... for a given set of known parental genotypes ?
- ... for hypothetical offspring comprising Sels, Full-sibs, Half-sibs and unrelated individuals ?

### [B] What are the attributes of a discriminant marker ?

### [C] Does the presence of null alleles affect the result ?

## RESULTS

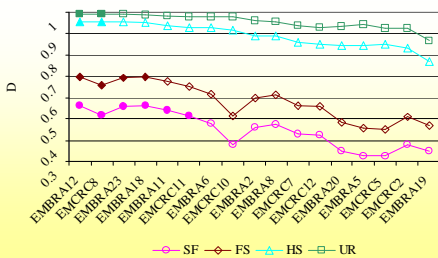
**Figure 1:** Discriminant power (D) and effective number of alleles (Ne) for the 17 SSR and 140 élites used in this study, with a 4 SSR MSDM (filled symbols).



**[A] MSDM**  
 4 SSR:  $C=2 \times 10^{-4}$   
 6 SSR:  $C=10^{-4}$   
 12 SSR:  $C < 10^{-6}$

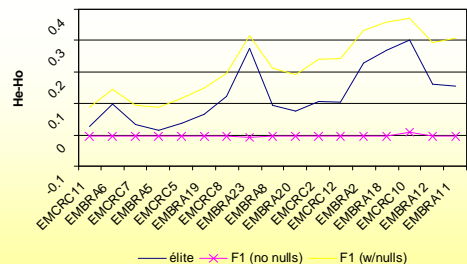
**[B] MSDM comprised systematically highly polymorphic SSR with  $N_e \geq 11$ , i.e. high polymorphism + low variance of allelic frequencies.**

**Figure 2:** Discriminant power (D), with MSDM (filled symbols) for simulated offspring, with pairs of unrelated individuals (UR; upper line), half-sibs (HS), full-sibs (FS) and pairs of sels (SF; lower line).



### [A] MSDM

6 SSR for SF:  $C=10^{-2}$   
 4 SSR for FS:  $C=10^{-2}$   
 2 SSR for HS:  $C=10^{-4}$   
 2 SSR for UR:  $C < 10^{-6}$

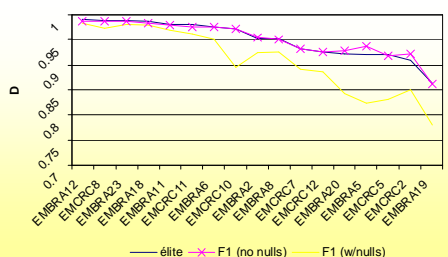


**Figure 3:** Difference between expected ( $H_e$ ) and observed heterozygosity ( $H_o$ ), for élites F0, simulated F1 from random mating of élites with no presence of null alleles, and simulated F1 from random mating of élites with null alleles (100% of all the homozygotes).

**[C]** The élite's deficit in H could be partly explained by the presence of null alleles. Note that this deficit followed a similar pattern to that shown by the F1 with null alleles.

**Figure 4:** D for same cases as in Fig. 3.

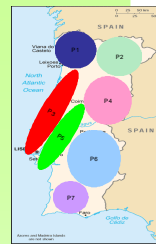
**[C]** For a limited number of scenarios, the presence of null alleles in the élites increased D in their simulated offspring (ex., case of parental carriers of 1 null allele). In general, however, null alleles decreased D in the offspring, especially for those SSR of low polymorphism.



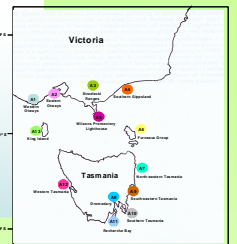
## MATERIAL AND METHODS

### RAIZ breeding Population (500 élites)

Sample of 140 élites and use of publicly available molecular markers (see ref.): 17 nuSSRs



RAIZ: 7 regions from P1 to P7 (seed introduction after 18th century)



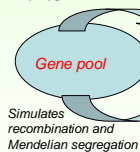
13 provenances

### Methodological approach

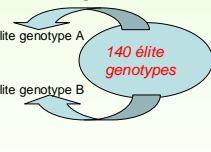
Development of ZETA<sup>®</sup> Monte-Carlo simulation software to obtain...

- ...basic genetic descriptors for each marker (heterozygosity, effective number of alleles, measures of relatedness, etc);
- ...MSDM with error = 0 (if enough markers) for the sample of 140 élites;
- ...MSDM with error > 0 for any hypothetical set of offspring from élites parents (or parental *gene pool*);
- ...and MSDM with the presence of null alleles.

For any hypothetical offspring from élites ...



For the sample of 140 élites...



Monte-Carlo D was obtained by bootstrap of  $10^{+6}$  pairs of genotypes: (i) of non-related, half-sibs, full-sibs and sels; (ii) existing elite genotypes.

**Discriminant power (D)** = proportion of 1's across bootstrapped cases.

Thus, the frequency of pairwise comparisons for which a marker presents distinguishable genotypic patterns over the assemble of comparisons across the sample under study.

The error of confusion (C) is equal to 1-D.

## CONCLUSIONS

**We presented a useful tool to analyse the discrimination power (D) and to minimize the subsequent risk of confusion (C) in the fingerprinting of genotyped breeding populations.**

**For the present case, safe fingerprinting was possible with a limited number of highly polymorphic SSR. The fingerprinting capacity of these SSRs was stable across sub-samples of distinct origin within the élite, and also between simulated offspring.**

**Fingerprinting capacity may be reduced by the presence of null alleles. The choice of highly polymorphic SSR could eventually minimize this effect.**

(\* ) publicly available user-friendly version of ZETA in preparation...

References : Brondani, R.P.V., Brondani, C., Tarchini, R. and Grattapaglia, D. (1998). *Theor Appl Genet* 97: 816–827. Brondani, R.P.V., Brondani, C., and Grattapaglia, D. (2002). *Mol Genet Genomics* 267: 338-347. Jones, R.C., Steane, D.A., Potts, B.M. and Vaillancourt, R. (2002). *Can J For Res* 32: 59-66. Steane, D.A., Vaillancourt, R.E., Russell, J., Powell, W., Marshall, D. and Potts, B.M. (2001). *Silvae Genetica* 50: 89-91.